

MARCIO FALEIROS VENDRAMINI

**VARIANTES NO GENE DA ADIPONECTINA
(ADIPOQ): RELAÇÕES COM ADIPONECTINEMIA
E DIABETES MELLITUS TIPO 2 EM
NIPO-BRASILEIROS**

Tese apresentada à Universidade
Federal de São Paulo – Escola Paulista
de Medicina para obtenção do Título de
Doutor em Ciências.

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Carlos, e às minhas queridas irmãs, Raquel e
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INTRODUÇÃO

O conceito clássico do tecido adiposo como um reservatório inerte de gordura mudou nos últimos quinze anos para um órgão endócrino ativo com funções regulatórias no metabolismo corporal, assim como na resposta imune e inflamatória. Essas ações são mediadas por peptídeos secretados pelo tecido adiposo que apresentam características em comum com as citocinas, sendo coletivamente referidos como adipoquinas ou adipocitoquinas, e compreendem, entre outros, a leptina, fator de necrose tumoral α (TNF- α) e adiponectina.

A adiponectina, identificada há aproximadamente dez anos atrás por quatro grupos independentes usando diferentes abordagens, é o peptídeo mais abundante secretado pelo tecido adiposo. Em 1995, Scherer e col., descreveram uma proteína de 30 kDa produzida exclusivamente no tecido adiposo que foi denominada ACRP30 (*adipocyte complement related protein of 30kDa*) devido a sua semelhança estrutural com a fração C1q do complemento⁽¹⁾. Essa proteína foi também identificada como o produto do DNAC isolado independentemente como adipoQ⁽²⁾ e APM1 (*AdiPose Most abundant Gene transcript 1*)⁽³⁾. No plasma humano, a proteína foi isolada como GBP28 (*gelatin binding protein 28*)⁽⁴⁾. Após a caracterização de sua organização genômica e localização no braço longo do cromossomo 3, a proteína foi denominada adiponectina⁽⁵⁾. Vários nomes foram dados a essa proteína, porém sua importância era desconhecida. Com o desenvolvimento de um método de ELISA (*enzyme-like immunoabsorbent assay*) para mensuração de seus níveis plasmáticos, revelou-se o significado clínico da adiponectina.

ESTRUTURA E MODO DE AÇÃO

O gene da adiponectina, atualmente denominado ADIPOQ de acordo com a nomenclatura da *Human Genome Organisation* (HUGO), localiza-se no cromossomo 3q27 e é composto por três *exons*. Nessa região identificou-se um *locus* de susceptibilidade para o Diabetes Mellitus tipo 2 em caucasianos e japoneses e também para traços quantitativos associados com a síndrome metabólica⁽⁶⁻⁸⁾. A adiponectina é uma proteína de 30 kDa e 244 aminoácidos em sua estrutura monomérica, sendo constituída por quatro domínios ou regiões: uma região amino-terminal de seqüência sinalizadora; uma região variável entre as espécies, que não apresenta homologia com outras proteínas; uma região colagenosa, que sofre glicosilação e hidroxilação; e uma seqüência carboxi-terminal, que forma o domínio globular altamente conservado entre as espécies e que apresenta homologia com o TNF- α ⁽⁹⁾(Figura 1).

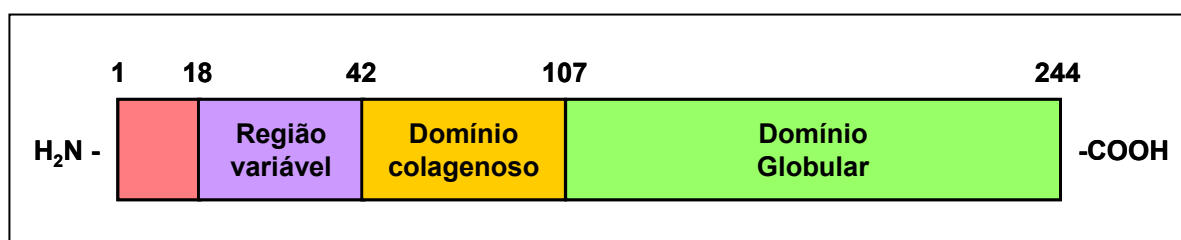


Figura 1: Estrutura primária da adiponectina humana.

Através de modificações pós-tradução de glicosilação e hidroxilação, a adiponectina apresenta-se em três principais formas oligoméricas: trímero de baixo peso molecular, hexâmero de peso molecular médio e multímero de alto peso molecular, contendo doze ou mais monômeros. Três monômeros se associam pela região globular para formar trímeros e esses, por sua vez,

multimerizam para formar hexâmeros e complexos de alto peso molecular⁽¹⁰⁾ (Figura 2). Aparentemente, essas interações envolvendo as regiões globular e colagenosa são importantes para a estabilidade e atividade das formas multiméricas. O resíduo cisteína na posição 36 (região variável amino-terminal) está envolvido na formação das pontes dissulfeto que são essenciais para a formação dos complexos oligoméricos; também a hidroxilação dos resíduos prolina e lisina nas regiões variável e colagenosa tem papel importante na ação insulino-sensibilizadora da adiponectina⁽¹¹⁾. A forma monomérica da adiponectina não é encontrada na circulação e parece estar confinada aos adipócitos. As isoformas presentes na circulação são estáveis e, aparentemente, não sofrem interconversão *in vivo*⁽¹²⁾. Estudos *in vitro* e *in vivo* indicam que a maior parte, se não a totalidade, da atividade biológica da adiponectina é atribuída aos complexos de alto peso molecular. Dados de estudos em seres humanos mostram que as concentrações dos multímeros de alto peso molecular refletem melhor as anormalidades metabólicas da obesidade e estão mais associadas ao desenvolvimento da síndrome metabólica do que as concentrações de adiponectina total⁽¹³⁻¹⁵⁾.

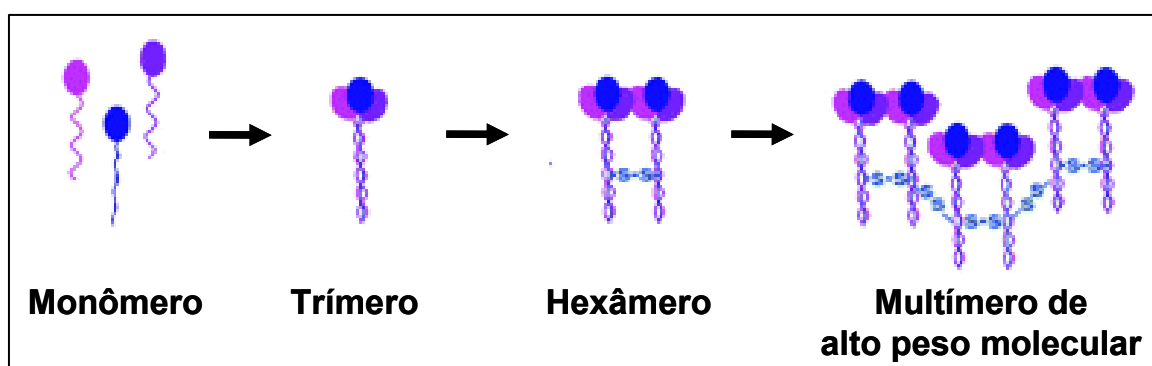


Figura 2: Formas multiméricas da adiponectina

Vários métodos têm sido utilizados para análise da distribuição das isoformas de adiponectina. Scherer e col. utilizaram técnicas de centrifugação em gradiente e *Western Blot* quantitativo para mensuração das diferentes formas de adiponectina *in vivo* e *in vitro*^(1,13). Tsao e col., empregaram cromatografia de filtração em gel para isolar os complexos de adiponectina^(16,17), enquanto Waki e col., desenvolveram um método mais simples, baseado em eletroforese em gel de poliacrilamida sob condições não redutoras e não denaturantes⁽¹⁸⁾. Recentemente, Nakano e col., desenvolveram um ELISA utilizando anticorpo monoclonal contra adiponectina humana de alto peso molecular⁽¹⁹⁾.

A adiponectina circula na corrente sanguínea em altas concentrações (3 a 30 µg/ml) em adultos saudáveis, correspondendo a aproximadamente 0,01% do total de proteína plasmática. Há um dimorfismo sexual nos seus níveis circulantes, com as mulheres apresentando valores mais elevados do que os homens^(20,21). Há ainda um dimorfismo sexual em relação aos complexos oligoméricos, com os homens apresentando uma menor proporção de multímeros de alto peso molecular em comparação com as mulheres⁽²²⁾.

Os efeitos pleiotrópicos da adiponectina são mediados por dois receptores de superfície clonados por Yamauchi e col., em 2003 e denominados AdipoR1 e AdipoR2. O AdipoR1 é expresso abundantemente na musculatura esquelética, enquanto que o AdipoR2 se expressa predominantemente no fígado. Ambos os receptores contêm sete domínios transmembrana, porém são estrutural e funcionalmente distintos dos receptores acoplados à proteína G. O AdipoR1 parece ter alta afinidade pela porção globular da adiponectina e baixa afinidade

para a proteína integral, enquanto que o AdipoR2 tem afinidade intermediária para ambas as formas⁽²³⁾.

Como principal produto secretor dos adipócitos, a adiponectina atua de maneira autócrina e parácrina dentro do tecido adiposo, e de maneira endócrina nos tecidos distais⁽²⁴⁾. Os efeitos autócrinos são ilustrados por seu papel na diferenciação dos adipócitos. Há indução maior do que 100 vezes do mRNA da adiponectina ao longo do curso da diferenciação das células adiposas⁽²⁵⁾. Igualmente, em experimentos envolvendo adipócitos que super-expressam adiponectina, a diferenciação celular foi acelerada, o que levou a aumento do acúmulo de lipídios e da atividade de transporte da glicose responsiva à insulina em células inteiramente diferenciadas. Assim, a adiponectina promove diferenciação dos adipócitos e aumento da sensibilidade à insulina^(25,26). Além disso, estudos recentes *in vivo* e *in vitro* forneceram evidências de uma alça de *feedback* reguladora, pela qual a adiponectina regula para baixo sua própria produção e a expressão de seu receptor⁽²⁷⁾. Além da capacidade de regular sua própria produção, a adiponectina atua como fator autócrino e parácrino para inibir a secreção pelos adipócitos de interleucinas 6 e 8, proteína-1 α/β inflamatória dos macrófagos e proteína-1 quimiotática dos monócitos, os quais, por sua vez, podem inibir o armazenamento de lipídios e a sensibilidade à insulina nos adipócitos^(24,28).

A adiponectina também pode atuar como fator endócrino. Foi demonstrado que a administração de adiponectina provoca um aumento na fosforilação insulino-estimulada da tirosina no receptor de insulina em músculo, tanto em roedores quanto em humanos^(29,30). Esse efeito pode contribuir para a

melhora da sensibilidade à insulina. Segundo Bacha e col., os níveis de adiponectina são responsáveis por 73% da variação na sensibilidade à insulina⁽³¹⁾.

FUNÇÕES BIOLÓGICAS DA ADIPONECTINA

Ação sensibilizadora à insulina

Estudos *in vitro* e *in vivo* demonstram uma forte correlação entre sensibilidade à insulina e níveis de adiponectina⁽³¹⁻³⁸⁾. Yamauchi e col. demonstraram melhora da sensibilidade à insulina e da hiperglicemia em modelos animais de obesidade, diabetes e lipoatrofia após a infusão sistêmica de doses fisiológicas de adiponectina⁽²⁹⁾. Além disso, estudos utilizando *clamp* euglicêmico hiperinsulinêmico mostram que o aumento agudo dos níveis de adiponectina circulantes através de infusão, melhora a supressão da produção hepática de glicose induzida pela insulina⁽³⁶⁾. Em estudo longitudinal e prospectivo, Hotta e col., demonstraram que os níveis circulantes de adiponectina decrescem paralelamente à progressão da resistência à insulina durante o desenvolvimento do Diabetes Mellitus (DM) tipo 2 em macacos Rhesus geneticamente predispostos à resistência à insulina⁽³⁵⁾. Nesses animais, a queda dos níveis de adiponectina precede o desenvolvimento da hiperglicemia. A correlação positiva mais forte observada nesse estudo foi com a taxa de captação de glicose estimulada pela insulina (valor M), um indicador de sensibilidade à insulina. Também em humanos, verificou-se que níveis diminuídos de adiponectina são um fator de risco para o desenvolvimento do DM tipo 2^(39,40).

Os efeitos sensibilizadores da insulina pela adiponectina parecem ser mediados pela fosforilação de AMPK e PPAR α no músculo esquelético e no fígado. No músculo esquelético, a ativação da AMPK leva ao aumento da oxidação de ácidos graxos livres e captação de glicose. A ativação do PPAR α pela adiponectina também estimula a oxidação de ácidos graxos e leva ao decréscimo do conteúdo de triglicérides no músculo, o que contribui para a melhora na transmissão do sinal insulínico⁽²⁹⁾. No fígado, a ativação da AMPK leva à redução de moléculas envolvidas na gliconeogênese e aumento da oxidação de ácidos graxos livres. A ativação do PPAR α também estimula a oxidação de ácidos graxos livres e a diminuição do conteúdo de triglicérides no fígado. Essas alterações promovem aumento da sensibilidade à insulina *in vivo*⁽⁴¹⁾.

Há dados que sugerem ainda um modo de ação central da adiponectina. A AMPK é envolvida na regulação hipotalâmica do comportamento alimentar e a adiponectina pode modular, via AMPK, o comportamento alimentar, provavelmente dentro da via da melanocortina⁽⁴²⁾.

Ação anti-aterosclerótica

A adiponectina tem efeitos biológicos nas células vasculares, incluindo células endoteliais, macrófagos e células da musculatura lisa.

A hipoadiponectinemia está associada com disfunção endotelial⁽⁴³⁾. Os efeitos da adiponectina nas células endoteliais são mediados pelo óxido nítrico, uma vez que ela reduz a supressão da atividade da óxido nítrico sintase (eNOS) induzida pela LDL oxidada e estimula a produção de óxido nítrico através

da via dependente da PI3K envolvendo a fosforilação da eNOS através da AMPK^(44,45).

Nos macrófagos, a adiponectina inibe a expressão do SRA-1 (*scavenger receptor class A-1*), resultando em diminuição da captação de LDL oxidada e inibição da formação de células espumosas⁽⁴⁶⁾. As células espumosas produzem vários tipos de metaloproteinases de matriz, as quais induzem ruptura das placas ateroscleróticas. A adiponectina aumenta a expressão e secreção de inibidores dessas metaloproteinases, via indução de IL-10, prevenindo, conseqüentemente, a ruptura dessas placas⁽⁴⁷⁾.

A adiponectina inibe a proliferação e migração de células da musculatura lisa vascular. Vários fatores ofensivos, como a LDL oxidada, estímulos inflamatórios e estresse oxidativo podem causar danos vasculares. Nessas situações, a adiponectina pode se acumular nas artérias danificadas e proteger contra o desenvolvimento da aterogênese^(48,49). Uma evidência *in vivo* do papel da adiponectina na aterosclerose é dada pela demonstração de que camundongos com *knockout* da adiponectina apresentam maior espessamento da neointima e aumento da proliferação de células da musculatura lisa em artérias submetidas a uma lesão mecânica. A suplementação de adiponectina a esse modelo animal atenua a proliferação da neointima⁽³⁷⁾.

Ação antiinflamatória

A inflamação tem um papel importante no desenvolvimento da aterosclerose. O fator de transcrição NFκB induz a expressão de citocinas e

moléculas de adesão no processo inflamatório. A adiponectina inibe a ativação do NFκB induzido pelo TNF-α sem afetar outros sinais mediados por ele⁽⁵⁰⁾.

VARIANTES NO GENE DA ADIPONECTINA

Baixos níveis de adiponectina têm sido associados com obesidade, resistência à insulina e aumento no risco de desenvolvimento de DM tipo 2 em diferentes estudos^(21,51-53). Além disso, estima-se que uma proporção substancial (30 – 70 %) da variabilidade dos níveis plasmáticos de adiponectina seja regulada por fatores genéticos⁽⁵⁴⁾. Assim sendo, é possível que variantes no gene ADIPOQ possam modular os níveis plasmáticos de adiponectina, bem como o risco para o desenvolvimento de resistência à insulina e DM tipo 2. De fato, variantes comuns e raras foram genotipadas em diferentes grupos étnicos e uma modulação genética para essas condições pôde ser demonstrada em diversos estudos⁽⁵⁵⁻⁵⁷⁾.

O gene ADIPOQ é muito polimórfico: vários polimorfismos foram encontrados nos *exons*, *introns* e regiões promotoras. Associações com adiponectinemia e/ou fenótipos relacionados à síndrome metabólica têm sido reportados para três grupos de variantes em várias populações: nas seqüências 5'(-12823, -11426, -11391 e -11377); na região do *exon 2* e *intron 2* (mutações +45 e +276) e no *exon 3* (G48R, G90S, Y111H, R112C e I164T)⁽⁵⁸⁾.

Kondo e col.,⁽⁵⁹⁾ fizeram o rastreamento de mutações no gene da adiponectina e verificaram que a frequência da mutação I164T foi significativamente mais elevada em portadores de DM tipo 2 quando comparada a controles pareados para idade e índice de massa corpórea. Além disso, as concentrações de adiponectina plasmática em indivíduos com a mutação I164T

foram inferiores às observadas em indivíduos sem a mutação e todos os indivíduos com a mutação apresentavam alguma característica da síndrome metabólica. Dois polimorfismos freqüentes, uma substituição silenciosa no *exon 2* (45T-G) e uma substituição de G por T no *intron 2* (276G-T) foram significativamente associados com DM tipo 2 em japoneses⁽⁵⁵⁾, com obesidade em alemães⁽⁵⁶⁾ e com obesidade e outros componentes da Síndrome Metabólica em não diabéticos italianos⁽⁵⁷⁾.

As mutações R112C, I164T, R221S e H241P foram detectadas exclusivamente em japoneses, enquanto que as mutações G90S, R92X e Y111H só foram encontradas em caucasianos. Apenas a rara mutação G84R foi encontrada nos dois grupos⁽⁶⁰⁾.

POPULAÇÃO NIPO-BRASILEIRA

O Brasil apresenta a maior população japonesa fora do Japão, provenientes de um movimento migratório que se estendeu de 1908 até o início da década de 1960, com uma interrupção de 1941 a 1952 (período da Segunda Guerra Mundial e pós-guerra imediato)⁽⁶¹⁾. Atualmente, há no Brasil cerca de 1,5 milhões de japoneses e descendentes, sendo que 80% residem no estado de São Paulo.

Estudos realizados na década de 1980 na população nikkei (segunda e terceira gerações) dos Estados Unidos mostram uma prevalência de DM três a quatro vezes superior àquela observada no Japão⁽⁶²⁾. Entretanto, estudos mais recentes mostram que essa diferença tende a diminuir, à medida que o Japão passa a incorporar hábitos ocidentais.

Diante da representatividade dos nipo-brasileiros na população brasileira e do impacto da imigração na saúde desses indivíduos, surgiu a idéia de aprofundar as investigações relativas à morbidade e mortalidade desse grupo populacional no Brasil. No final da década de 1980, constituiu-se então, no Departamento de Medicina Preventiva da Escola Paulista de Medicina, o grupo multiprofissional de pesquisadores para estudos sobre a população nipo-brasileira coordenado, até seu falecimento, pelo Prof. Dr. Magid Lunes e, desde então, pela Profa. Dra. Sandra Roberta G. Ferreira, e denominado *Japanese-Brazilian Diabetes Study Group* (JBDSG) para fins de divulgação de trabalhos no meio científico⁽⁶¹⁾.

Inicialmente, foram aplicados questionários bilíngües na cidade de São Paulo com a intenção de avaliar a prevalência de DM auto-referido entre japoneses e seus descendentes. Notou-se que, de fato, a prevalência era maior do que aquela encontrada na população brasileira em geral e na do Japão, sendo similar à encontrada entre nipo-americanos de Seattle⁽⁶³⁾. No entanto, a necessidade de se avaliar a real prevalência de DM entre nipo-brasileiros, acabou levando à procura de um local com número representativo de membros da comunidade e que contasse com facilidade de acesso ao centro de coleta e oferta de recursos humanos e de infra-estrutura para a realização de exames médicos e laboratoriais. Após visitas a inúmeras instituições em diferentes municípios, optou-se pela cidade de Bauru, no interior do estado de São Paulo. A partir daí, definiu-se como população-alvo aquela formada por indivíduos de origem japonesa, de primeira e segunda gerações, não-miscigenados e com idade entre 40 e 79 anos⁽⁶¹⁾.

O projeto de estudo do diabetes e doenças associadas iniciou-se em 1992 com o levantamento da população nikkei de Bauru, então formada por cerca de 3000 indivíduos entre 40 e 79 anos. Todos os indivíduos de primeira geração e um terço dos de segunda geração, de ambos os sexos, foram convidados a participar da primeira fase de estudos (1993). No total, 647 indivíduos foram examinados e a prevalência de DM foi de 22,6%, superior àquela observada na população geral brasileira e entre os que viviam no Japão⁽⁶¹⁾.

A segunda fase do estudo (2000) foi mais abrangente, analisando todos os indivíduos de primeira e segunda gerações, a partir dos 30 anos de idade. O número de casos novos de DM ocorridos nesse período foi alarmante, e a prevalência da doença já atingia 36,1%, uma das maiores já registradas em todo o mundo⁽⁶⁴⁾.

A possibilidade que se aventa é que os nipo-brasileiros, geneticamente predispostos ao desenvolvimento de resistência à insulina, quando expostos a um ambiente desfavorável apresentam uma série de alterações metabólicas, entre elas, o DM tipo 2. Além da identificação de fatores de risco ambientais, a identificação de fatores de risco genético permitirá adoção precoce de medidas preventivas e possibilidade de um tratamento mais adequado, com conseqüentes implicações prognósticas.

OBJETIVOS

Tendo em vista os aspectos abordados acima, os objetivos do presente estudo foram:

- Investigar se baixos níveis plasmáticos de adiponectina constitui fator de risco independente para o desenvolvimento de intolerância à glicose na população nipo-brasileira (Artigo 1).
- Investigar a frequência, significância clínica e a contribuição para a ocorrência de DM em nipo-brasileiros dos polimorfismos T45G, G276T e A349G, e das mutações no *exon 3* do gene ADIPOQ (Artigo 2).
- Reportar uma nova mutação no gene da adiponectina associada com hipoadiponectinemia em nipo-brasileiros (Artigo 3).

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Plasma adiponectin levels and incident glucose intolerance in Japanese–Brazilians: A seven-year follow-up study

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ABSTRACT

The objective of this study was to investigate whether decreased baseline adiponectin levels are an independent risk factor for development of glucose intolerance in a population-based study of Japanese–Brazilians, a group with one of the highest prevalence rates of diabetes worldwide. We examined 210 Japanese–Brazilians (97 male and 113 female, aged 56.7 ± 10.1 years) with normal glucose tolerance (NGT). Plasma adiponectin, insulin, fasting and 2-h plasma glucose and lipid profile were evaluated at baseline and also at 7-year follow-up. Plasma adiponectin levels were significantly lower in glucose intolerance progressors compared with subjects who remained NGT. By increasing tertiles of adiponectin, the frequencies of subjects who progressed to glucose intolerance were 40%, 33% and 27% and the frequencies of subjects who remained NGT were 13%, 35% and 52% ($\chi^2 = 15.8$, $p = 0.001$). Logistic regression analyses showed that adiponectin levels (OR for the highest versus lowest tertile: 0.31; 95% CI: 0.12–0.84, $p = 0.021$), male sex (OR: 2.61, 95% CI: 1.21–5.65, $p = 0.015$), fasting plasma glucose (OR: 3.05, 95% CI: 1.35–6.91, $p = 0.008$) and waist circumference (OR: 1.04, 95% CI: 1.00–1.08, $p = 0.046$) were independent risk factors for the progression to glucose intolerance. In conclusion, low plasma levels of adiponectin is one of several independent predictors of glucose intolerance in a Japanese–Brazilian population.

1. Introduction

Diabetes mellitus is a serious and costly disease that profoundly impacts health and longevity. Thus, it may be more efficient to identify individuals at risk in order to provide measures to prevent the progression into diabetes.

Adiponectin, the gene product of the most abundant gene transcript 1 (apM1), is a collagen-like protein that is exclusively expressed and secreted by the adipose tissue⁽¹⁾. Recent data have demonstrated a strong correlation between plasma adiponectin and insulin sensitivity both *in vivo* and *in vitro* studies^(2,3). Decreased concentrations are seen in states of insulin resistance, including type 2 diabetes and obesity, while improvement of insulin sensitivity by weight loss or administration of thiazolidinediones has been shown to increase plasma adiponectin concentrations⁽⁴⁻⁷⁾. Furthermore, circulating adiponectin levels have been shown to decrease in parallel with the progression of insulin resistance and to precede diabetes development in rhesus monkeys genetically predisposed to insulin resistance⁽²⁾. The predictive value of low adiponectin for future development of type 2 diabetes in humans has been previously demonstrated in a few population groups⁽⁸⁻¹²⁾. Japanese–Brazilians show a high prevalence of glucose metabolism disturbances and incidence rates of diabetes mellitus, being the prevalence rates of diabetes one of the highest worldwide⁽¹³⁾. In this population, after 7-year follow-up, we observed an overall high proportion of subjects (approximately 75%) in all categories of glucose intolerance. However, a greater increase was observed in the category of isolated impaired fasting glucose (from 3.3% to 19.3%). Also, a gradual and statistically significant increase in mortality rates was observed as fasting or 2-h plasma glucose elevated⁽¹⁴⁾. In this study, as part of an 7-year follow-up of Japanese–Brazilians, we examine the importance of low levels of plasma adiponectin as a predictor for the development of glucose intolerance in this population.

2. Materials and Methods

2.1. Study population

In 1993 we started to follow-up a sample of subjects from the Japanese–Brazilian community living in Bauru, a developed city of the São Paulo State, to investigate the prevalence of type 2 diabetes and its associated risk factors. This study involved all individuals aged 40–79 years from the first generation (Issei) and a random sample of one-third (with additional 20%) of those from the second generation (Nisei) from the same age-group. The entire population of second generation was listed in alphabetical order and every third subject was selected to compose the Nisei group studied. The percentage of refusal to participate was 11.8% for Issei and 11.1% for Nisei. Six hundred and forty-seven first-generation (37.3%) and second-generation (62.7%) Japanese–Brazilians aged 40–79 years went through an interview with questionnaires about demographic and social aspects, medical history and nutritional survey, assessed by food frequency questionnaire. Clinical examination was performed including anthropometric (weight, height and waist circumference) and blood pressure measurements. Fasting blood samples for glucose, insulin, lipids and adiponectin were obtained. A 75 g solution of anhydrous glucose was administered and 2 h later another blood sample was obtained for glucose and insulin measurements. Seven years later (from 1999 to 2000) the glucose tolerance status of 394 subjects was reexamined (follow-up rate: 61%) and the same parameters evaluated at baseline were evaluated again. Of the original study sample, 69 subjects (10.6%) had died, 57 (8.7%) had moved and 127 (19.7%) refused to participate. For this study we enrolled 210 individuals (97 male and 113 female, aged 56.7 ± 10.1 years) with normal glucose tolerance according to the 1993 OGTT and suitable baseline plasma specimens.

2.2. Laboratory methods

Plasma glucose was determined by the glucose-oxidase method. Cholesterol contents of lipoproteins fractions and triglycerides were measured enzymatically. Insulin was determined by a monoclonal antibody-based immunofluorimetric assay⁽¹⁵⁾. Insulin resistance was estimated by indirect measures which included fasting insulin concentrations and homeostasis model assessment (HOMA-IR)⁽¹⁶⁾. Glucose tolerance status was based on 1999 WHO criteria⁽¹⁷⁾. At the 7-year follow-up 52 subjects had normal glucose tolerance (NGT), 42 had impaired fasting glucose (IFG), 73 had impaired glucose tolerance (IGT) and 43 had diabetes (DM). Since the categories of IFG and IGT are not mutually exclusive, subjects meeting criteria for both categories were classified as IGT. Fasting plasma adiponectin concentrations were measured by radioimmunoassay (Linco Research, St. Charles, MI, USA) in baseline plasma specimens that had been stored at -20°C . This assay has a sensitivity of 1 ng/ml with an intra- and inter-assay coefficients of variation of 1.8–6.2% and 6.9–9.3%, respectively.

This study was approved by the Ethics Committee of Escola Paulista de Medicina, Universidade Federal de São Paulo and all participants were informed about the aims of the study and gave their consent.

2.3. Statistical analysis

All data are shown as mean \pm SD. Differences in continuous variables among the subgroups were evaluated by unpaired Student's t-test. Data were tested for normal distribution with Kolmogorov–Smirnov test and non-normally distributed parameters were logarithmically transformed to approximate normal distribution. The non-parametric Mann–Whitney test was used on parameters which were still not normally distributed after transformation. Baseline adiponectin concentrations were divided into tertiles (≤ 5.27 ; 5.28–9.90 and ≥ 9.91 $\mu\text{g/ml}$) and the relationships between adiponectin and development of glucose intolerance were evaluated by χ^2 analyses. In order to identify independent associations

between development of glucose intolerance by the 7-year follow-up examinations and parameters collected at baseline, logistic regression was used. An odds ratio for each tertile of adiponectin was estimated. Variables were entered into the model due to a p -value < 0.20 in univariate analyses. Potential interactions between the covariates were calculated, but were excluded from the model due to $p > 0.05$.

3. Results

At the 7-year follow-up a total of 158 of 210 individuals, who had normal glucose tolerance at baseline (75.2%, 95% CI: 68.8–80.9%) developed glucose intolerance (IGT, IFG or DM). The comparison of subjects' baseline characteristics showed a worse metabolic profile among those who progressed to glucose intolerance (Table 1). Subjects who remained NGT had lower mean values of BMI, waist circumference, fasting and 2-h plasma glucose concentrations, triglycerides and higher mean values of HDL than those who progressed to glucose intolerance. Also, plasma adiponectin levels were significantly higher in subjects who remained NGT compared with the glucose intolerance progressors. Plasma adiponectin concentrations were significantly higher in women ($11.01 \pm 7.29 \mu\text{g/ml}$) than in men ($6.97 \pm 4.59 \mu\text{g/ml}$; $p < 0.001$) and there was a higher percentage of men who progressed to glucose intolerance compared to women (87.6% versus 64.6%, $p < 0.001$). Next, we examined the relationships between glucose intolerance development and adiponectin levels. Baseline adiponectin levels were divided in tertiles and the frequency of subjects with NGT or glucose intolerance by the 7-year follow-up examination is shown in Fig. 1. By increasing tertiles of adiponectin, the frequencies of subjects who progressed to glucose intolerance fell progressively being 40%, 33% and 27% whereas the frequencies of subjects who remained NGT rose, being 13%, 35% and 52% ($\chi^2 = 15.8$, $p = 0.001$).

In order to identify risk factors for the progression to glucose intolerance in this population, we performed a multiple logistic regression. Sex, waist circumference, fasting plasma glucose, diastolic blood pressure, HDL cholesterol,

triglycerides and adiponectin levels were used as covariates. Considering the possibility of multicollinearity between waist circumference and BMI, BMI was excluded from the multivariate analyses and waist circumference was included as representative of trait related to obesity. Table 2 shows that adiponectin level is an independent factor for developing glucose intolerance. The odds ratios by increasing tertiles of adiponectin were 1.0, 0.33 (95% CI: 0.12–0.90) and 0.31 (95% CI: 0.12–0.84). Male sex and fasting plasma glucose were also independent risk factors for the progression to glucose intolerance, as was waist circumference (Table 2).

4. Discussion

Type 2 diabetes, a worldwide growing health problem, has insulin resistance as a fundamental element. But, despite of intensive investigations little is known about the factors responsible for insulin resistance in persons at increased risk of developing the disease.

In this prospective study of Japanese–Brazilians with normal glucose tolerance we examined the relationships between baseline adiponectin concentrations and the development of glucose intolerance. At the 7-year follow-up 75.2% of individuals developed glucose intolerance. This is a very high rate of conversion, showing that diabetes mellitus and its long-term complications should be considered as one of the major public health problems in this population. Our data also emphasize the need of primary prevention programs in this community. There is some evidence of ethnic variations in adiponectin concentrations^(18,19). Indeed, plasma adiponectin levels found in Japanese–Brazilians were similar to those in native Japanese, since both populations are thought to be genetically identical. We found higher adiponectin levels in women than in men. Even after adjusting for possible confounding factors (data not shown), gender remained an independent predictor of serum adiponectin levels. This observation is in agreement with previous studies in humans⁽²⁰⁻²³⁾ and in mice⁽²⁴⁾.

We demonstrated that low baseline levels of adiponectin were significantly associated with the risk of developing glucose intolerance over 7 years of follow-

up. This association was independent of sex, age, waist circumference, as a measure of visceral obesity and other modifiers of diabetes risk. Subjects in the lowest tertile of adiponectin developed glucose intolerance 3.2 times (95% CI: 1.2–8.3) more than those in the highest tertile ($p = 0.021$). Adiponectin may exert its protective effect in the development of glucose intolerance by improving insulin action and secretion. Weyer et al.,⁽²⁵⁾ reported in Caucasian and Pima Indians that adiponectin levels are closely related to the degree of insulin resistance. Other investigators have presented data confirming this connection⁽²⁰⁻²⁶⁾. Also, decreased tyrosine phosphorylation of muscle insulin receptor was associated to low plasma adiponectin concentrations⁽²⁶⁾. Recently, Rakatzi et al.,⁽²⁷⁾ identified a novel function of adiponectin consisting of the inhibition of cytokine and fatty acid induced apoptosis and dysfunction of pancreatic beta cells. Previous studies demonstrated the predictive value of low adiponectin levels for future development of type 2 diabetes in various ethnic groups such as Pima Indians, Asian Indians, Japanese, African Americans and Caucasians^(8,9,11,12). However, our study showed a predictive value of low adiponectin levels for all categories of glucose intolerance (IFG, IGT and DM).

These data have potential implications for pathophysiology, screening and prevention of glucose metabolism disturbances. Low levels of adiponectin in apparently healthy subjects may help to identify high-risk individuals for glucose intolerance. Therefore, preventive measures adopted early may reduce the prevalence of these disturbances. It was shown previously that weight reduction therapy increased plasma adiponectin concentration in nondiabetic and diabetic subjects and in severely obese subjects following bariatric surgery^(4,23). Also, the administration of thiazolidinediones increased adiponectin levels in non-diabetic and type 2 diabetic subjects⁽⁵⁻⁷⁾. However, it remains to be determined whether an increase of adiponectin levels per se will lead to a reduction in risk of glucose metabolism disturbances.

A limitation of the present study is the relatively low follow-up rate. Of the original study sample, 383 individuals had NGT at baseline and for this study 210 of them were enrolled. However, no difference in baseline characteristics such as age, BMI, waist circumference, sex distribution and fasting plasma glucose levels

(data not shown) was found between individuals enrolled and non-enrolled. Therefore, it probably did not cause any bias.

In summary, we found that low plasma levels of adiponectin were an independent predictor over 7 years to glucose intolerance in a subset of Japanese–Brazilian subjects with normal baseline glucose tolerance. It is suggested that adiponectin may have a role in the mechanisms of glucose intolerance development. However, a relatively raised fasting blood glucose within the normal range remains the strongest predictor of future type 2 diabetes in this population.

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Table 1: Baseline characteristics of the subjects who remained NGT and the ones who progressed to glucose intolerance at 7-year follow-up.

	NGT	Glucose intolerance	<i>p</i>
Age (years)	57.4 ± 11.2	56.4 ± 9.8	0.511 ^a
BMI (kg/m²)	22.9 ± 3.2	24.8 ± 3.7	0.001 ^a
Waist (cm)	79.9 ± 11.0	86.1 ± 9.6	<0.001 ^a
Men	83.1 ± 7.8	87.5 ± 8.6	0.101 ^a
Women	79.0 ± 11.7	84.4 ± 10.4	0.012 ^a
Fasting plasma glucose (mmol/l)	4.8 ± 0.4	5.1 ± 0.5	0.001 ^a
2 h plasma glucose (mmol/l)	5.1 ± 0.9	5.5 ± 1.2	0.006 ^b
Fasting insulin (pmol/l)	27.5 ± 27.9	33.7 ± 57.3	0.764 ^c
Systolic blood pressure (mmHg)	123.5 ± 17.1	127.0 ± 18.1	0.228 ^a
Diastolic blood pressure (mmHg)	76.0 ± 10.0	78.7 ± 10.7	0.114 ^a
Cholesterol (mmol/l)	5.5 ± 1.0	5.5 ± 1.3	0.937 ^a
HDL (mmol/l)	1.2 ± 0.3	1.1 ± 0.3	0.004 ^a
Triglycerides (mmol/l)	1.7 ± 2.0	1.8 ± 1.2	0.038 ^c
Adiponectin (µg/ml)	12.2 ± 7.5	8.0 ± 5.6	<0.001 ^d
HOMA-IR	5.9 ± 6.0	7.7 ± 12.6	0.975 ^c

Data are means ± S.D.

^a *t*-Test.

^b *t*-Test to unequal variance.

^c Mann–Whitney test.

^d *t*-Test performed on log transformations Data are means ± S.D.

Table 2: Multiple logistic model for identification of prognostic factors to glucose intolerance progression.

	Odds ratio	95% CI	p
Sex (M/F)	2.61	[1.21–5.65]	0.015
Adiponectin (5.28–9.90 vs. 0.85–5.27)	0.33	[0.12–0.90]	0.030
Adiponectin (>9.90 vs. 0.85–5.27)	0.31	[0.12–0.84]	0.021
Fasting plasma glucose (mmol/l)	3.05	[1.35–6.91]	0.008
Waist (cm)	1.04	[1.00–1.08]	0.046

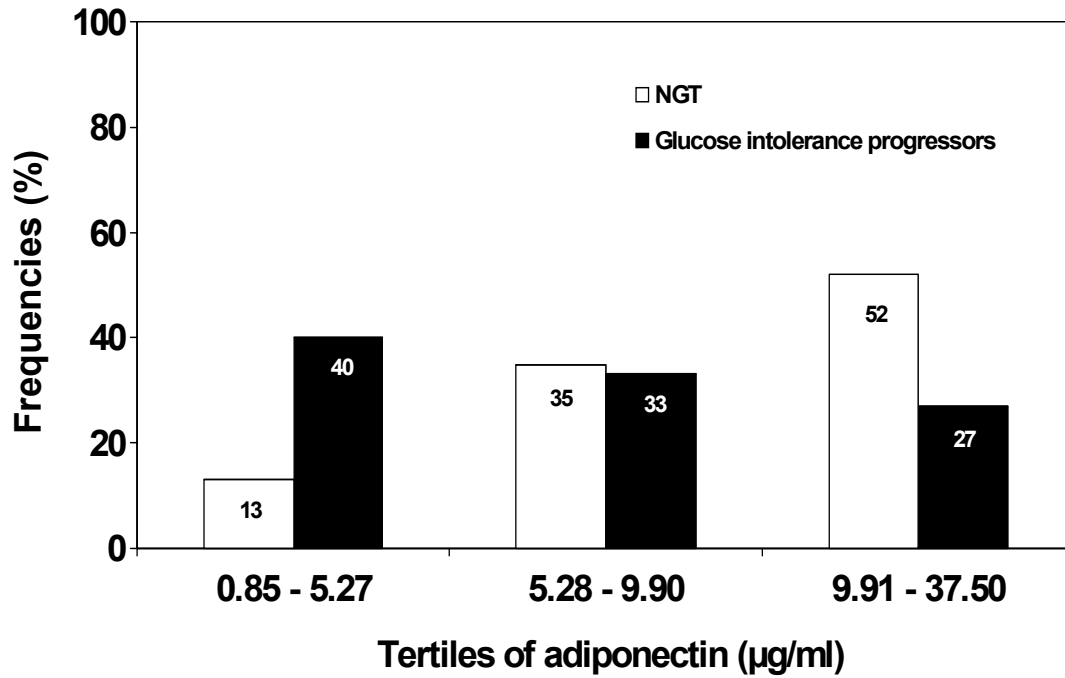


Figure 1: Distribution of subjects who remained NGT and those who progressed to glucose intolerance at 7-year follow-up according to tertiles of baseline adiponectin.

Association of genetic variants in the adiponectin encoding gene (ADIPOQ) with type 2 diabetes in Japanese-Brazilians

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ABSTRACT

Aim: To assess the contribution of ADIPOQ variants to type 2 diabetes in Japanese-Brazilians.

Methods: we genotyped 200 patients with diabetes mellitus (100 male and 100 female, aged 55.5 ± 10.8 years) and 200 control subjects with NGT (72 male and 128 female, aged 54.0 ± 13.4 years).

Results: whereas each polymorphism studied (T45G, G276T and A349G) was not significantly associated with type 2 diabetes mellitus, the haplotype GGA was over-represented in our diabetic population (9.3% against 3.1% in NGT individuals, $p = 0.0003$). Also, this haplotype was associated with decreased levels of adiponectin. We also identified three mutations in exon 3: I164T, R221S and H241P but, owing the low frequencies of them, associations with type 2 diabetes could not be evaluated. The subjects carrying the R221S mutation had plasma adiponectin levels lower than those without the mutation (1.97 ± 0.80 $\mu\text{g/ml}$ vs 8.08 ± 6.27 $\mu\text{g/ml}$, $p = 0.015$). Similarly, the I164T mutation carriers had mean plasma adiponectin levels lower than those non-carriers (3.73 ± 0.88 $\mu\text{g/ml}$ vs 8.08 ± 6.27 $\mu\text{g/ml}$) but this difference was not significant ($p = 0.23$).

Conclusions: we identified in the ADIPOQ gene a risk haplotype for type 2 diabetes that affects plasma adiponectin levels in the Japanese-Brazilian population.

Key words: adiponectin, Japanese-Brazilian, single nucleotide polymorphisms, ADIPOQ gene

1. Introduction:

Adiponectin, an adipose tissue-specific protein, is secreted and exists in the circulation of healthy individuals at high concentrations, accounting for approximately 0.01% of total plasma protein. It plays an important role in modulating insulin sensitivity and hypoadiponectinemia is seen in obesity, type 2 diabetes and coronary artery disease, conditions closely related to insulin resistance (Arita et al., 1999; Hotta et al., 2000; Weyer et al., 2001). Adiponectin is encoded by the ADIPOQ gene located on chromosome 3q27, a region with evidence for linkage with type 2 diabetes and the metabolic syndrome. The ADIPOQ gene spans 16 kb, contains three exons (translation starts at exon 2 and ends at exon 3) (Takahashi et al., 2000; Schaffler et al., 2000). Frequent and rare variants of ADIPOQ gene were genotyped in large data sets from various ethnics groups. The variants T45G and G276T are the most common and have been the most widely studied, both separately and as haplotypes. These variants have been associated with type 2 diabetes in some studies (Hara et al., 2002; Stumvoll et al., 2002; Menzaghi et al., 2002). However, depending on the population studied results have been inconsistent. Less common point mutations located in exon 3 were also identified. The I164T mutation, detected exclusively in the Japanese population, was associated with type 2 diabetes, metabolic syndrome and coronary artery disease (Kondo et al., 2002; Ohashi et al., 2004).

The Japanese-Brazilians show a high prevalence of glucose metabolism disturbances and incidence rate of diabetes mellitus, being the prevalence rates of diabetes one of the highest worldwide (Gimeno et al., 2002). We previously described that low plasma concentrations of adiponectin is one independent predictor of glucose intolerance in this population (Vendramini et al., 2006). In the present study, we investigated the contribution of SNPs T45G, G276T and A349G of the ADIPOQ gene to diabetes in the Japanese-Brazilian population. Also, the frequency and clinical significance of less common point mutations in exon 3 were examined.

2. Subjects and Methods:

2.1. Study subjects

Mutation screening was carried out on genomic DNA from 200 randomly selected patients with diabetes mellitus (100 male and 100 female, aged 55.5 ± 10.8 years) and 200 age-matched control subjects with normal glucose tolerance (72 male and 128 female, aged 54.0 ± 13.4 years). All the individuals were recruited from the Japanese-Brazilian Diabetes Study Group, a survey designed to estimate the prevalence and incidence of diabetes and associated diseases in a Japanese-Brazilian population living in Bauru, São Paulo, Brazil. Details on the selection and recruitment of the sample population have been previously described (Gimeno et al., 2002). All individuals went through an interview with questionnaires about demographic and social aspects and medical history. Clinical examination was performed including anthropometric (weight, height and waist circumference) and blood pressure measurements. Fasting blood samples for glucose, insulin, lipids and adiponectin were obtained. A 75g solution of anhydrous glucose was administered and two hours later another blood sample was obtained for glucose and insulin measurements. This study was approved by the Ethics Committee of Escola Paulista de Medicina, Universidade Federal de São Paulo, and all participants were informed about the aims of the study and gave their consent.

2.2. Methods:

Plasma glucose was determined by the glucose-oxidase method. Cholesterol contents of lipoproteins fractions and triglycerides were measured enzymatically. Insulin was determined by a monoclonal antibody-based immunofluorimetric assay (PerkinElmer, Wallac Oy, Turku, Finland). Insulin resistance was estimated by indirect measures which included fasting insulin concentrations and homeostasis model assessment (HOMA-IR) (Matthews et al., 1985). Glucose tolerance status was based on 1999 WHO criteria (Alberti et al., 1999). Fasting plasma adiponectin concentrations were measured by radioimmunoassay (Linco Research, St Charles, MI, USA). This assay has a sensitivity of 1 ng/ml with an intra and interassay coefficients of variation of 1.8-6.2% and 6.9-9.3%, respectively.

Blood samples were obtained from each subject and genomic DNA was extracted from peripheral blood leukocytes using a commercial kit (Puregene DNA Isolation Kit, Genra System, Minneapolis, MN, USA). The entire translated region of adiponectin gene was amplified by PCR using two pairs of specific primers: 5'AGAAAGCAGCTCCTAGAAGT3' and 5'GGCACCATCTACACTCATCC3' which flank the region containing exon 2, and 5'CCTAAGGGAGACATCGGTGA3' and 5'ATTGACTTTGGGGCTGTTTG3' which flank the region containing exon 3. The PCR products were directly sequenced on an ABI Prism 3100 Genetic Analyzer (Applied Biosystems, CA, USA). Numbering of nucleotides: the A of the ATG of the initiator Met codon was denoted as +1 (Gen Bank accession number D45371.1) and amino acids were numbered according to Gen Bank accession number NP_004788.

2.3. Statistical analysis:

For continuous variables, data are given as mean \pm SE. The chi-square test was used to compare the genotype or allele frequencies between study groups. Differences in continuous variables between the two groups were calculated by the Student t test. Haplotype frequency, Hardy-Weinberg equilibrium and linkage disequilibrium statistics were obtained through the use of Haploview software. Association analysis for alleles and haplotypes for the dichotomic variable diabetes was conducted using Haploview. Association analysis for genotypes were conducted using SPSS ver. 13.0. Association analysis for alleles and haplotypes for adiponectin levels were conducted using the Hapstat software. A p value $< 0,05$ was considered significant.

3. Results:

The clinical characteristics of type 2 diabetes patients and control subjects are shown in Table 1. Patients with type 2 diabetes had significantly higher BMI and HOMA-IR index and lower plasma adiponectin levels than control subjects. The genotype and allele frequencies of SNPs T45G, G276T and A349G are shown in Table 2. No significant allele or genotype association could be disclosed for the diabetes phenotype, although allele 349G tended to be less prevalent in diabetic individuals (unadjusted p-value 0.06). Adjusted p-values (gender, age and

BMI) for allelic effects also did not reach statistical significance for the diabetic phenotype. There was no evidence of Hardy-Weinberg disequilibrium for any SNP.

On Figure 1 we present the linkage disequilibrium relationship between tested markers. Considering the linkage disequilibrium structure of this particular locus we have proceeded to test whether any haplotype could be associated with an increased risk of presenting diabetes. We analyzed the 4 haplotypes with frequency > 5%: TGA: 0.411; TTA: 0.278; GGG: 0.224 and GGA: 0.062. A positive association was observed with haplotype GGA (45G + 276G + 349A), that was over-represented in our diabetic population (9.3% in type 2 diabetic individuals against 3.1% in normal glucose tolerant individuals, $p = 0.0003$). On the contrary, the GGG haplotype was more frequent in normal glucose tolerant individuals as compared to diabetic individuals (25.5% versus 19.4%, respectively, $p = 0.04$). Both associations remained statistically significant even after model adjustment for age (data not shown). Haplotypes were not independently associated with BMI levels.

We next examined the genotype influence on adiponectin levels. None of studied alleles showed significant association with adiponectin levels (Table 3). However, a significant haplotype effect in adiponectin levels was disclosed for the GGA haplotype (Table 4). In accordance to what would be expected, the estimated effect of the GGA haplotype is to decrease adiponectin levels.

We also investigated the influence of less common nonsynonymous point mutations located in exon 3. In the present study we identified three mutations, previously detected in the Japanese population: I164T, R221S and H241P. All the subjects with the mutations were heterozygous. The I164T mutation was found only in two diabetic subjects. The R221S was identified in two diabetic subjects and in one normal glucose tolerant individual. The H241P mutation was identified in three diabetic subjects and in four normal glucose tolerant subjects. Owing the low frequencies of these mutations, the association between them and type 2 diabetes could not be evaluated. The subjects carrying the R221S mutation had plasma adiponectin levels lower than those without the mutation ($1.97 \pm 0.80 \mu\text{g/ml}$ vs $8.08 \pm 6.27 \mu\text{g/ml}$, $p = 0.015$). Similarly, the I164T mutation carriers had mean plasma adiponectin levels lower than those non-carriers ($3.73 \pm 0.88 \mu\text{g/ml}$ vs $8.08 \pm 6.27 \mu\text{g/ml}$) but this difference was not significant ($p = 0.23$).

4. Discussion

Some previous studies have shown that ADIPOQ gene variants are associated with type 2 diabetes in cross-sectional as well as prospective studies (Hara et al., 2002; Stumvoll et al., 2002; Menzaghi et al., 2002; Vasseur et al., 2003; Kondo et al., 2002; Ohashi et al., 2004). However, these associations have not been seen in all studies and the contribution of each variant in different populations is not established.

A silent T to G substitution in exon 2 (T45G) and a G to T substitution in intron 2 (G276T) have been the most widely studied variants, both separately and as haplotypes. In the Japanese population, Hara et al. found that subjects with the genotype GG at the position 45 or the genotype GG at position 276 had an increase risk of type 2 diabetes (Hara et al., 2002). Also, in Chinese people, the T45G was the only polymorphism significantly associated with the risk of persistent hyperglycemia at 5-year follow-up and haplotypes formed by the addition of other SNPs did not confer greater association (Tso et al., 2006). However, Gu et al studying Swedish Caucasians found no significant differences of allele frequencies for SNPs 45 and 276 between type 2 diabetic patients and non diabetic controls (Gu et al., 2004). Similarly, in the Nurses' Health Study polymorphisms at positions +45 and +276 were not associated with risk of diabetes and the haplotype defined by these two SNPs was also not associated with diabetes risk (Hu et al., 2004). Here, studying Japanese-Brazilians, a population characterized by a high prevalence of glucose intolerance, we provided evidence that a haplotype defined by SNPs 45, 276 and 349 (GGA) contributed to the genetic risk of type 2 diabetes.

In order to test whether the effects of haplotype were due to effects on adiponectin levels, plasma adiponectin was measured in all of the subjects examined. In accordance to the observed association with type 2 diabetes, the GGA haplotype was associated with decreased levels of adiponectin. The biology of this association is unclear. One possibility is that the GGA haplotype may act decreasing adiponectin expression; however none of the three SNPs are functional. The T45G is in exon 2 and is a silent polymorphism and G276T and A349G are in intron 2 where no regulatory sites have been described. Thus, this

haplotype is more likely to be in linkage disequilibrium with one or more functional variants affecting plasma adiponectin levels.

In addition to common polymorphisms, rare nonsynonymous mutations located in exon 3 have been described in ADIPOQ gene (Kondo et al., 2002). In the present study we identified the I164T, R221S and H241P mutations previously reported in the Japanese population. The I164 mutation was found only in two subjects, both with type 2 diabetes. The two subjects mutation-carriers, besides diabetes, had at least one more metabolic disorder such as hypertension and dyslipidemia. Mean plasma adiponectin levels in these subjects were lower than in wild-type, but this difference was not statistically significant possibly because of the small number of patients with the mutation. This finding is in agreement with previous studies in Japanese population showing a correlation between I164T mutation and type 2 diabetes (Kondo et al., 2002) and lower plasma adiponectin levels (Kondo et al., 2002; Ohashi et al., 2004). Investigating the mechanisms by which this mutation cause diabetes and hypoadiponectinemia, Waki et al. (2003) showed that the I164T mutants are not assembled into stable trimers and larger multimers, which probably resulted in impaired secretion from the cell. However, Kishida et al. (2003) found an oligomerization state similar to the wild-type, but abnormal secretion from adipose tissue.

Regarding the R221S mutation, Kondo et al. (2002) identified only one nondiabetic subject carrying this mutation, with adiponectin levels similar to the non-carriers. We found three mutation carriers with adiponectin levels lower than the non-carriers. However, more individuals carrying the R221S mutation need to be identified for better assessment of the association between this mutation and adiponectin levels.

In summary, we identified in the ADIPOQ gene a risk haplotype for type 2 diabetes that affects plasma adiponectin levels in the Japanese-Brazilian population.

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Table 1: Clinical characteristics of normal glucose tolerant (NGT) subjects and type 2 diabetic patients.

	NGT Subjects (n= 200)	Type 2 Diabetic Patients (n= 200)	p value
Age (years)	54.0 ± 13.4	55.5 ± 10.8	0.23
Sex (M/F)	72 / 125	100 / 100	0.0092
BMI (kg/m²)	22.8 ± 2.75	27.4 ± 3.72	< 0.0001
HOMA IR	1.20 ± 0.51	4.89 ± 3.42	< 0.0001
Adiponectin (µg/ml)	10.62 ± 6.74	6.48 ± 5.02	< 0.0001
Systolic BP (mmHg)	123.8 ± 22.1	138.3 ± 24.4	< 0.001
Diastolic BP (mmHg)	76.0 ± 12.6	82.9 ± 15.4	< 0.001
Triglycerides (mg/dL)	235.7 ± 197.0	325.8 ± 278.4	< 0.0001

Table 2: Genotype and allele frequencies of SNPs T45G, G276T and A349G in NGT subjects and type 2 diabetic patients (T2DM).

SNP	Genotypes n(%)				Alleles n(%)		
	T/T	T/G	G/G		T	G	
+45							
T2DM	93 (46.5)	95 (47.5)	12 (6.0)	0.61	281 (70.2)	119 (29.8)	0.74
NGT	99 (50.2)	84 (42.6)	14 (7.1)		282 (71.6)	112 (28.4)	
+276							
T2DM	85 (42.5)	104 (52.0)	11 (5.5)	0.25	274 (68.5)	126 (31.5)	0.18
NGT	97 (49.2)	94 (47.7)	6 (3.0)		288 (75.0)	106 (25.0)	
+349							
T2DM	118 (61.8)	64 (33.5)	9 (4.7)	0.16	300 (78.5)	82 (21.5)	0.06
NGT	100 (50.8)	80 (40.60)	12 (6.1)		280 (72.9)	104 (27.1)	

Table 3: Effect estimative of studied alleles on adiponectin levels.

	Effect estimative (p value)
T45 allele	+ 0.16 (0.75)
G276 allele	+ 0.65 (0.23)
A349 allele	- 0.58 (0.28)

Models adjusted for age.

Effect estimative - change in mean adiponectin levels according to allele groups.

Table 4: Effect estimative of studied haplotypes on adiponectin levels.

	Effect estimative (p value)
TGA	- 0.40 (0.37)
TTA	+ 0.75 (0.18)
GGG	+ 0.47 (0.37)
GGA	- 1.90 (0.04)

Models adjusted for age.

Effect estimative - change in mean adiponectin levels according to haplotypes groups.

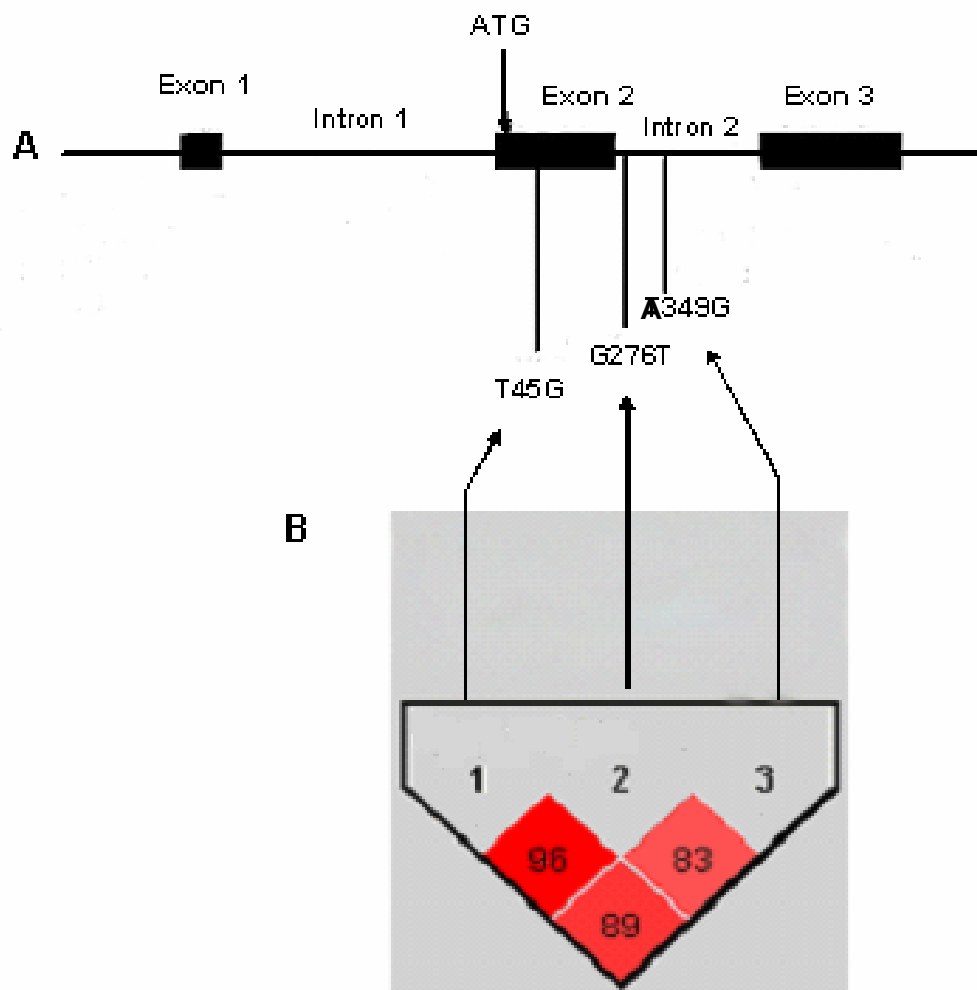


Figure 1. Schematic representation of *ADIPOQ* gene and linkage disequilibrium (LD) structure.

A. The exon-intron organization of the gene is indicated by boxes and lines, the positions of studied variants are showed.

B. Relative LD between SNP pairs is indicated by the scheme, based on the absolute values of the Lewontin normalized coefficient multiplied by 100, defining a sole haplotype block.

A Novel Mutation in the Adiponectin (ADIPOQ) Gene is Associated with Hypoadiponectinemia in Japanese-Brazilians

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Running title: novel mutation in ADIPOQ gene and hypoadiponectinemia

Key words: adiponectin, ADIPOQ gene, Japanese-Brazilians

Abstract

Objective: Adiponectin, an important mediator of insulin sensitivity, is encoded by the *ADIPOQ* gene. Here we describe two Japanese-Brazilian families with hypoadiponectinemia due to a novel mutation on *ADIPOQ* gene.

Design and Patients: In this study, we examined the entire translated regions of adiponectin in Japanese-Brazilians, a population with one of the highest prevalence rates of diabetes worldwide. We screened 200 patients with type 2 diabetes and 240 age-matched subjects with normal glucose tolerance.

Results: A novel heterozygous T deletion at position 186 in exon 2 of the *ADIPOQ* gene, causing a frameshift at codon 62 leading to a premature termination at codon 168 (*p.Gly63ValfsX106*) was found in two individuals with diabetes. This mutation was not found in 240 nondiabetic control subjects. In addition, we screened the mutation in an expanded set of 100 nondiabetic subjects from general Brazilian population, but we found no mutation. Six additional mutation carriers family members of probands were identified. Mutation-carrier individuals had markedly low plasma adiponectin concentrations compared with those without the mutation (DM: 1.01 ± 0.69 $\mu\text{g/ml}$ vs 6.49 ± 5.04 $\mu\text{g/ml}$, $p < 0.001$; NGT: 1.10 ± 0.49 $\mu\text{g/ml}$ vs 10.36 ± 6.63 $\mu\text{g/ml}$, $p = 0.003$). All individuals carrying the *p.Gly63ValfsX106* mutation and older than 30 years were found to be diabetic.

Conclusions: We describe for the first time a frameshift mutation in exon 2 of the *ADIPOQ* gene, which modulates adiponectin levels and may contribute to the genetic risk of late-onset diabetes in Japanese-Brazilians.

Introduction

Adiponectin is an adipocyte-secreted protein that has been recognized as an important mediator of insulin sensitivity (1, 2). Thus, adiponectin may play a role in the pathogenesis of type 2 diabetes, obesity and atherosclerotic cardiovascular disease, conditions closely related to insulin resistance. In fact, several studies have reported low plasma adiponectin levels in patients with these diseases (3-5). It circulates as a multimeric high-order structures comprised of homotrimers that associates through disulphide bonds to form low molecular weight (LMW) hexamers (~180 kDa) and high molecular weight (HMW) multimers of > 300 kDa. Pajvani et al. showed that the ratio, and not the absolute amounts, between HMW to total adiponectin correlates better with measures of insulin sensitivity, suggesting that the HMW adiponectin complex is the active form of this protein (6).

Mutational screening of the *ADIPOQ*, the gene encoding adiponectin, was performed in different ethnic groups and an association of genetic variants with type 2 diabetes and insulin resistance was found (7-11).

Japanese-Brazilians show a high prevalence of glucose metabolism disturbances, being the prevalence rates of diabetes one of the highest worldwide (12). In this population we found that low plasma levels of adiponectin were an independent predictor over 7 years to glucose intolerance in a subset of subjects with normal glucose tolerance at baseline (13).

In the present study, we examined the entire translated regions of adiponectin gene in Japanese-Brazilians and a novel mutation associated with hypoadiponectinemia is reported.

Subjects and Methods

Subjects: The mutation screening was carried out on genomic DNA from 200 randomly selected patients with diabetes mellitus (100 male and 100 female, aged 55.4 ± 10.7 years) and 240 age-matched control subjects with normal glucose tolerance (81 male and 159 female, aged 53.9 ± 13.0 years). All the individuals were recruited from the Japanese-Brazilian Diabetes Study Group, a survey designed to estimate the prevalence and incidence of diabetes and associated diseases in a Japanese-Brazilian population living in Bauru, São Paulo, Brazil. Details on the selection and recruitment of the sample population have been previously described (12). All individuals went through an interview with questionnaires about demographic, social aspects and medical history. Clinical examination was performed including anthropometric (weight, height and waist circumference) and blood pressure measurements. Fasting blood samples for glucose, insulin, lipids and adiponectin were obtained. A 75 g solution of anhydrous glucose was administered and two hours later another blood sample was obtained for glucose and insulin measurements. This study was approved by the Ethics Committee of Escola Paulista de Medicina, Universidade Federal de São Paulo, and all participants were informed about the aims of the study and gave their consent.

Biochemical measurements: Plasma glucose was determined by the glucose-oxidase method. Cholesterol contents of lipoproteins fractions and triglycerides were measured enzymatically. Insulin was determined by a monoclonal antibody-based immunofluorimetric assay (PerkinElmer, Wallac Oy, Turku, Finland). Insulin resistance was estimated by homeostasis model assessment (HOMA-IR) (14). Glucose tolerance status was based on 1999 WHO criteria (15). Fasting plasma

adiponectin concentrations were measured by radioimmunoassay (Linco Research, St Charles, MI, USA).

Mutation screening: Blood samples were obtained from each subject and genomic DNA was extracted from peripheral blood leukocytes using a commercial kit (Puregene DNA Isolation Kit, Gentra System, Minneapolis, MN, USA). The entire translated region of adiponectin gene was amplified by PCR using two pairs of specific primers: 5'AGAAAGCAGCTCCTAGAAGT3' and 5'GGCACCATCTACACTCATCC3' which flank the region containing exon 2, and 5'CCTAAGGGAGACATCGGTGA3' and 5'ATTGACTTTGGGGCTGTTTG3' which flank the region containing exon 3. The PCR products were directly sequenced on an ABI Prism 3100 Genetic Analyzer (Applied Biosystems, CA, USA). Sequence changes of frameshift mutation were determined by cloning the PCR product using the TOPO TA Cloning (Invitrogen, Carlsbad, CA, USA) and sequencing clones derived from both alleles. Numbering of nucleotides: +1 of ATG codon (GenBank: D45371.1) and amino acids according to GenBank accession number NP_004788.

SDS-PAGE and immunoblotting for determination of adiponectin oligomers:

SDS-PAGE was performed according to standard Laemmli procedure. Human serum diluted 10 fold was solubilized in 5x Sample buffer and loaded on a 3-12 % polyacrilamide gel under non-reducing, nonheated-denaturing conditions. Proteins were then electrophoretically transferred into a nitrocellulose membrane (Hybond ECL, Amersham Biosciences, USA). After blocking for 1 hour in 5% nonfat milk in TBS with 0,05% Tween 20, the blots were incubated with a human monoclonal antibody from BD Bioscience (1:5000 dilution with 1% nonfat milk in TTBS) for 2 h at room temperature. After washings, the membrane was incubated with

horseradish peroxidase-conjugated anti-rabbit antibody (1:5000 dilution) (Santa Cruz Biotechnology, USA) for 1 h at room temperature and then washed thoroughly. Bands were detected using ECL Western blotting detection reagent (Amersham Biosciences, USA) and quantified by densitometry analyses using the Scion Image software. Relative proportions of adiponectin oligomers were calculated by dividing band density by total density in each lane. This analysis was performed in all diabetic mutation-carrier individuals (n=5) and in five diabetic non mutation-carriers paired for age, gender and BMI.

Microsatellite studies: three polymorphic microsatellite near the transcriptional unit of adiponectin gene were genotyped in two individuals carrying the p.Gly63ValfsX106 mutation from different families.

Statistical analysis. All data are shown as mean \pm SD. Differences in continuous variables among the groups with and without the mutation were evaluated by unpaired Student's t test or Mann-Whitney, when appropriate. Differences in the frequencies of mutation between groups were tested by χ^2 test. A p-value of < 0.05 was considered statistically significant.

Results

The main characteristics of the study population are shown in Table 1. Patients with diabetes had significantly higher BMI, HOMA-IR, waist circumference and lower adiponectin levels than the normal glucose tolerant subjects. By direct sequencing of 2 exons of the *ADIPOQ* gene we identified a novel heterozygous T deletion at position 186 in exon 2, causing a frameshift at codon 62 leading to a premature termination at codon 168 (*c.186delT*; *p.Gly63ValfsX106*, according to Human Genome Variation Society nomenclature) in two individuals with diabetes.

This mutation was not found in the 240 age-matched nondiabetic control subjects. In addition, we screened the mutation in an expanded set of 100 nondiabetic subjects from general Brazilian population (32 male, 66 female, aged 48.9 ± 14.4 years), but we found no mutation. Available family members of probands with the p.Gly63ValfsX106 mutation were genotyped for this variant (n= 14) and 6 additional carriers were identified, three of them with diabetes (Figure 1).

The diabetic patients carrying the p.Gly63ValfsX106 mutation had markedly low plasma adiponectin concentration compared with those diabetic patients without the mutation ($1.01 \pm 0.69 \mu\text{g/ml}$ vs $6.49 \pm 5.04 \mu\text{g/ml}$, $p < 0.001$). Similarly, between the individuals with normal glucose tolerance, those carrying the mutation had a statistically significant lower plasma adiponectin concentrations than the non-carriers ($1.10 \pm 0.49 \mu\text{g/ml}$ vs $10.36 \pm 6.63 \mu\text{g/ml}$, $p = 0.003$). This difference was not only limited to the absolute amount of adiponectin but also the ratio of HMW to total adiponectin was decreased among diabetic subjects carrying the mutation when compared to age, gender, BMI paired diabetic subjects without the mutation (0.17 ± 0.08 vs 0.32 ± 0.06 , respectively, $p = 0.018$) (Figure 2). In order to examine if hypoadiponectinemia is a familial issue not related to the p.Gly63ValfsX106 mutation, we compared adiponectin levels in mutation-carrier and non-mutation carrier individuals from F1 family. We found lower adiponectin levels in carriers than in non-carriers ($0.85 \pm 0.41 \mu\text{g/ml}$ vs $5.15 \pm 2.3 \mu\text{g/ml}$, $p = 0.009$).

Among the p.Gly63ValfsX106 carriers, we found that the diabetics were older (55.8 ± 4.71 years vs 21.7 ± 6.66 years, $p = 0.0001$) and had higher waist circumference (89.0 ± 7.24 cm vs 73.0 ± 9.54 cm, $p = 0.035$) than the normal glucose tolerant carriers. Also, the average age of diagnosis of diabetes was higher than

the chronological age of nondiabetic carriers (47.2±7.78 years vs 21.7±6.66 years, $p=0.003$) (Table 2).

Adiponectin levels showed a negative correlation with HOMA-IR ($r=-0.255$, $p<0.0005$) in the total Japanese-Brazilian study group. However, HOMA-IR values were not different in mutation-carriers compared to non-mutation carriers (Table 2).

Microsatellite studies showed different haplotypes for the three polymorphic markers in the two patients from F1 and F2 families, indicating that these two families have different genetic backgrounds. We, therefore, can infer that the mutations arose independently.

Discussion

Here we report for the first time a frameshift mutation with a premature termination in the *ADIPOQ* gene associated with hypoadiponectinemia in two families of Japanese-Brazilians. These two families come from distinct regions of Japan (Hokkaido and Chugoku) and the haplotype analysis derived from three polymorphic microsatellites indicated that these families have different genetic backgrounds. Therefore, a common origin or founder effect is unlikely. The search for the *p.GlyValfsX106* mutation in other populations will reveal if it is restricted to the Japanese Brazilian population. This variant was not identified in 680 alleles of normal subjects, ruling out the possibility of polymorphism.

The *p.Gly63ValfsX106* mutation causes a frameshift at codon 62 leading to a premature termination at codon 168. As result, more than half of globular domain is missed. This mutation is located in the highly conserved collagen domain, which contains four lysines residues at positions 65, 68, 77 and 101 each having the

surrounding motif of GXKGE(D), that are also highly conserved between species. These four lysines were found to be hydroxylated and subsequently glycosylated. Wang et al. showed that in murine adiponectin the substitution of these four conserved lysines with arginine produced a variant with impaired ability to enhance insulin action in hepatocytes (16). Recently, Richards et al. (17) showed that in human adiponectin the conservative substitution of different combinations of these lysines has a more or less severe effect on the efficiency of HMW formation, demonstrating an important functional role of these lysine residues in the intracellular assembly and secretion of HMW multimers. Thus, the *p.Gly63ValfsX106* mutation, which leads to a substitution of two of these lysines and their surrounding motifs, may interfere with the post-translational modifications that regulate the multimer composition of adiponectin. In fact, when we assessed the multimer composition of adiponectin in serum by immunoblot and densitometric analysis we found a relative decrease of HMW forms in subjects carrying the mutation when compared to gender, age and BMI paired subjects without the mutation.

The gold standard for measuring insulin sensitivity is the hyperinsulinemic-euglycemic clamp technique, which is invasive, expensive and time-consuming to perform. Although not being the gold standard, HOMA-IR has been widely used as a tool to assess insulin sensitivity in clinical research and epidemiologic studies. In the present study we found a negative correlation between HOMA-IR and adiponectin levels in the total Japanese-Brazilian study group. However, despite of mutation-carriers being characterized by hypoadiponectinemia their HOMA-IR values were not different from non-mutation carriers. It should be pointed that since HOMA-IR is not a particularly precise measure of insulin, it is more suitable

for studies in large samples, such as the analyze in the total study group, than for assessment of individuals.

All individuals carrying the *p.Gly63ValfsX106* mutation and older than 30 years were found to be diabetic. The three normal glucose tolerant mutation-carriers individuals were younger (14, 25 and 26 years old) and may develop diabetes at later age. Based on the fact that in Japanese-Brazilians hypoadiponectinemia is an independent risk factor for the occurrence of glucose intolerance (13), these data suggests that heterozigosity for this mutation and the consequent hypoadiponectinemia may be a risk factor for late-onset diabetes.

Missense mutations in human adiponectin have been reported and some of them were associated with diabetes and hypoadiponectinemia (7, 8, 11). Investigating the mechanisms by which these mutations caused diabetes and hypoadiponectinemia, Waki et al found that mutants with impaired multimerization were associated with diabetes or hypoadiponectinemia while mutants without overtly abnormal phenotype showed normal multimerization (18). These data suggested impaired multimerization and/or consequent impaired secretion might be the causes of these phenotypes.

In conclusion, we describe for the first time a frameshift mutation in exon 2 of the *ADIPOQ* gene which modulates adiponectin levels and may contribute to the genetic risk of late-onset diabetes in Japanese-Brazilians.

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Table 1: Characteristics of study population by glucose tolerance status.

	DM (n=200)	TGN (n=240)	p value
Age (years)	55.4 ± 10.6	53.9 ± 13.0	NS
Sex (M/F)	100/100	81/159	< 0.001
BMI (kg/m²)	27.4 ± 3.7	23.1 ± 2.9	< 0.001
HOMA-IR	4.91 ± 3.4	1.20 ± 0.5	< 0.001
Waist circumf. (cm)	90.0 ± 11.3	78.2 ± 8.5	< 0.001
Adiponectin (µg/ml)	6.44 ± 5.0	10.36 ± 6.6	< 0.001

Table 2: Clinical and biochemical characteristics of type 2 diabetic patients and normal glucose tolerance subjects who bear the new mutation.

Subject	Age (years)	Sex (M/F)	BMI (kg/m ²)	Age at onset of DM (years)	Waist circumference (cm)	HDL-c (mg/dl)	Triglycerides (mg/dl)	HOMA-IR	Adiponectin (µg/ml)	
DM with mutation	F1 II-1	63	M	26.0	49	100.0	44	351	2.65	0.65
	F1 II-4	56	M	22.9	55	85.0	75	759	1.25	0.60
	F1 II-9	50	F	20.7	34	81.0	49	212	3.13	0.55
	F2 II-1	56	M	23.9	50	87.5	39	182	6.87	2.20
	F2 II-2	54	F	28.2	48	91.5	44	186	8.10	1.05
DM without mutation (n=198)	55.4 ± 10.7	98M 100F	27.5 ± 3.7	53.6 ± 11.3	89.9 ± 9.6	49.0 ± 11.9	326 ± 279.6	4.91 ± 3.4	6.49 ± 5.0	
p value	0.93	0.99	0.06	0.21	0.85	0.83	0.77	0.74	< 0.001	
NGT with mutation	F1 III-1	26	M	26.3	----	84	53	70	1,06	1.65
	F1 III-2	25	M	20.6	----	67	51	60	0.82	0.7
	F1 III-5	14	M	22.3	----	68	37	74	1.27	0.95
NGT without mutation (n=240)	53.9 ± 13.0	81M 159F	23.0 ± 2.9	----	78.1 ± 8.5	50.8 ± 10.5	108.8 ± 127.9	1.17 ± 0.5	10.3 ± 6.6	
p value	0.003	0.07	0.99	----	----	0.62	0.007	0.72	0.03	

Figure legends:

Figure 1: Pedigree of the families with *p.Gly63ValfsX106* mutation. The individuals with diabetes mellitus are represented with filled symbols, the subjects with normal glucose tolerance with empty symbols, and the subjects with impaired glucose tolerance with dashed symbols. Arrows indicate the probands. The text below each individual represents: age (years), BMI (kg/m^2), and serum adiponectin levels ($\mu\text{g}/\text{ml}$); M: mutant allele; N: wild-type allele; nd: not done.

Figure 2: Representative immunoblot illustrating the three multimer species of adiponectin from serum of diabetic patients mutation carrier (NM) and non-mutation carrier (NN). HMW: high molecular weight, MMW: medium molecular weight, LMW: low molecular weight

Figure 1

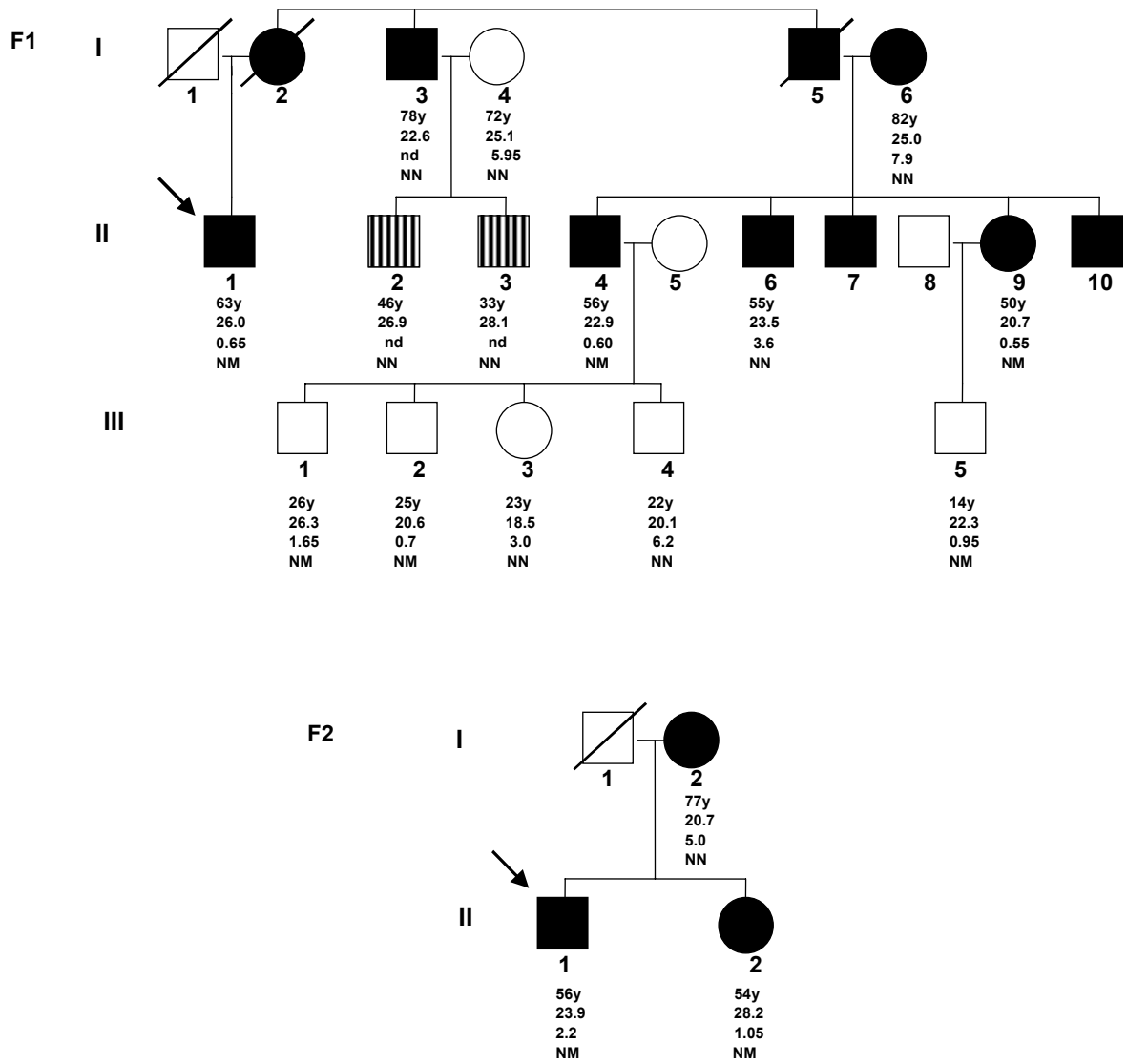
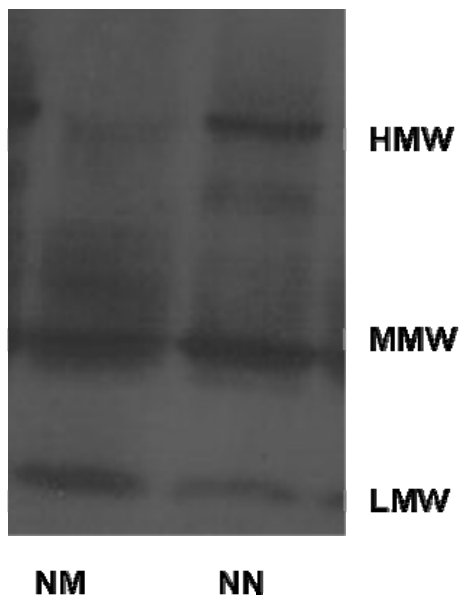


Figure 2



CONSIDERAÇÕES FINAIS

O tecido adiposo, além reservatório de energia, tem papel importante na regulação da homeostase energética, sensibilidade à insulina e metabolismo dos carboidratos e lipídios. Essas ações são mediadas através da secreção de vários peptídeos bioativos, referidos como adipocitoquinas.

Dentre as adipocitoquinas tem merecido destaque a adiponectina, que circula em concentrações relativamente altas no plasma, por suas importantes ações metabólicas e antiinflamatórias. Diferentes estudos mostram que os níveis circulantes de adiponectina são mais baixos em indivíduos obesos, portadores de Diabetes Mellitus (DM) tipo 2 e doença arterial coronariana, condições essas relacionadas à resistência à insulina. Portanto, níveis circulantes de adiponectina são índices de sensibilidade à insulina, porém ainda havia controvérsia se a adiponectinemia seria um simples marcador ou teria algum papel causal na sensibilidade à insulina. Estudos longitudinais em macacos mostraram que uma queda na adiponectinemia precede o desenvolvimento de resistência à insulina e DM tipo 2. Em humanos, estudos prospectivos avaliaram o papel do decréscimo nos níveis de adiponectina como preditor de DM ou síndrome metabólica. Ainda, estima-se que uma proporção substancial (30–70%) da variabilidade dos níveis plasmáticos de adiponectina seja regulada por fatores genéticos. Assim sendo, é possível que variantes no gene ADIPOQ possam modular os níveis plasmáticos de adiponectina, bem como o risco para o desenvolvimento de resistência à insulina e DM tipo 2.

Em nosso primeiro artigo, avaliamos a importância de baixos níveis de adiponectina como fator preditor para o desenvolvimento de intolerância à glicose na população nipo-brasileira, população essa caracterizada por alta prevalência de DM. Demonstramos que, em indivíduos com tolerância à glicose normal, níveis mais baixos de adiponectina estavam significativamente associados a um maior risco de desenvolvimento de intolerância à glicose em um período de sete anos de seguimento. Baixos níveis de adiponectina podem, portanto, ajudar a identificar entre indivíduos saudáveis aqueles com maior risco de desenvolvimento de intolerância à glicose, permitindo a instituição precoce de medidas preventivas.

No segundo artigo aqui apresentado, investigamos a contribuição das variantes comuns T45G, G276T e A349G no gene ADIPOQ para a ocorrência de DM na população nipo-brasileira. Examinamos também a frequência e a significância clínica das mutações no *exon 3* desse gene. Verificamos que o haplótipo GGA (45G + 276G + 349A) contribui para o risco genético do DM e é associado com níveis mais baixos de adiponectina. Além disso, detectamos a presença das mutações I164T, R221S e H241P, previamente descritas apenas na população japonesa, bastante raras na nossa população.

No terceiro artigo, ainda estudando o gene ADIPOQ, descrevemos pela primeira vez, em duas famílias nipo-brasileiras não relacionadas, uma mutação *frameshift* em heterozigose no *exon 2*, associada a níveis bastante reduzidos de adiponectina plasmática. A mutação *pGly63ValfsX106* leva a uma proteína truncada, com perda de parte significativa da porção globular, o que pode interferir na formação da adiponectina de alto peso molecular, considerada

biologicamente ativa. De fato, notamos um decréscimo relativo da forma de alto peso molecular entre os indivíduos carreadores da mutação, quando comparados a não carreadores pareados para sexo, idade e índice de massa corpórea. Portanto, verificamos que a modulação dos níveis de adiponectina e resultante maior suscetibilidade ao DM tipo 2 são causados tanto por variantes freqüentes quanto raras.

CONCLUSÕES

Os dados obtidos no presente estudo nos permitem concluir que:

- Baixos níveis plasmáticos de adiponectina foram preditores independentes para o desenvolvimento intolerância à glicose em nipo-brasileiros em um período de seguimento de sete anos (Artigo 1).
- Identificamos no gene ADIPOQ um haplótipo de risco para o DM tipo 2, que afeta os níveis plasmáticos de adiponectina em nipo-brasileiros (Artigo 2).
- Descrevemos pela primeira vez uma mutação *frameshift* no exon 2 do gene ADIPOQ (*p.Gly63ValfsX106*), a qual modula os níveis de adiponectina e pode contribuir para o risco genético de DM tipo 2 em nipo-brasileiros (Artigo 3).

Tabela 1: Características demográficas, antropométricas e laboratoriais dos indivíduos com TGN no período basal (Artigo 1).

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m²)	Adiponectina (µg/ml)
1	TM	M	61	99	47	78,5	20,7	11,67
2	HSY	F	66	260	53	82,5	23,5	4,55
3	NO	M	63	110	42	101,8	29,3	6,14
4	KH	F	62	169	44	88,0	23,3	29,44
5	MH	M	68	141	43	83,0	23,6	5,4
6	TY	M	46	342	36	108,0	34	1,45
7	AK	M	56	96	39	87,0	26,6	4,65
8	ES	F	63	109	49	78,3	22,5	8,56
9	ASK	M	52	112	38	79,0	23,1	6,48
10	BKK	M	60	115	40	84,5	23,6	4,99
11	HI	F	72	91	57	81,0	23,5	8,59
12	AAYA	F	50	264	49	84,8	25,3	10,85
13	FU	F	56	287	30	102,0	29,7	6,59
14	FH	M	59	151	47	80,5	21,3	11,33
15	MSY	F	63	58	55	88,5	23,1	11,34
16	FO	F	57	282	26	93,8	25,1	9,9
17	CHM	M	41	292	38	99,0	27,3	5,75
18	HN	F	53	39	69	63,5	17,5	22,49
19	AM	F	53	210	44	91,8	29,1	5,24
20	AST	M	44	259	35	88,0	25,6	4,5
21	AK	M	47	181	53	92,0	27,3	2,42
22	NNM	F	52	42	47	84,8	28,2	12,91
23	YH	M	56	121	29	73,5	19,8	2,23
24	KK	M	50	333	22	88,0	24,2	13,95
25	MMM	F	41	181	35	76,5	24,2	12,84
26	KM	F	70	134	37	100,3	28,1	5,3
27	HM	F	58	46	75	61,8	16,5	28,8
28	SU	M	60	234	45	96,0	28,7	7,42
29	KT	M	59	105	40	88,0	27,3	4,87
30	HN	F	60	122	68	54,6	14,9	14,2

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m²)	Adiponectina (µg/ml)
31	EF	M	69	95	42	82,5	23,2	14,31
32	M	44	98	44	72,0	19,7	6	M
33	M	68	101	45	84,0	22,3	9,18	M
34	M	50	405	29	87,0	19,6	2,23	M
35	F	60	57	59	74,0	18,9	12,45	F
36	M	48	146	38	85,5	23,3	6,5	M
37	M	61	133	48	76,0	18,9	8,72	M
38	F	43	89	91	57,0	17,4	23,49	F
39	M	60	181	41	90,3	25,1	9,59	M
40	F	62	161	39	87,5	23,2	19,13	F
41	F	58	207	34	88,0	25,0	21,88	F
42	M	45	280	34	79,0	20,9	1,96	M
43	M	56	104	55	108,3	31,6	2,17	M
44	M	56	54	51	96,3	24,6	16,21	M
45	F	44	86	38	76,8	25,0	4,69	F
46	F	69	146	48	75,3	18,1	22,27	F
47	F	72	157	36	72,8	21,6	5,64	F
48	F	59	139	40	88,3	24,1	5,75	F
49	M	47	171	46	70,3	21,8	6,38	M
50	F	68	275	39	88,8	25,7	8,5	F
51	M	52	491	31	95,5	30,0	4,12	M
52	F	64	153	42	95,5	26,8	11,81	F
53	M	64	79	41	75,5	19,5	2,33	M
54	M	65	90	61	81,8	21,0	9,07	M
55	F	57	120	45	91,0	20,5	5,05	F
56	M	67	149	38	89,5	25,1	5,11	M
57	F	50	42	50	67,0	21,7	30,31	F
58	F	68	192	37	81,5	24,9	3,05	F
59	M	57	85	56	75,5	21,2	17,51	M
60	M	71	156	30	93,5	28,1	6,66	M

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m²)	Adiponectina (µg/ml)
61	TT	M	68	199	50	88,5	25,1	5,26
62	KM	M	55	142	63	72,5	20,8	8,05
63	KOO	F	47	40	63	86,0	23,1	14,2
64	TT	F	63	88	51	73,0	22,4	17,8
65	NS	M	54	872		100,5	27,1	13,92
66	IG	M	54	168	32	85,5	23,9	4,36
67	LU	M	52	117	55	93,8	24,4	6,53
68	KKT	F	45	55	43	80,0	21,4	13,44
69	LKO	M	61	283	28	87,5	22,9	5,53
70	IT	M	56	89	38	89,3	27,1	5,3
71	HB	F	75	163	45	84,5	22,4	9,35
72	KK	M	50	128	31	111,5	35,5	3,91
73	MI	F	61	238	38	91,0	26,4	8,22
74	HH	M	42	350	33	81,5	33,1	4,29
75	JO	F	58	111	53	99,5	30,0	13,92
76	SMK	F	54	83	54	75,5	19,0	6,81
77	NTM	M	50	173	41	86,0	23,9	2,71
78	NNM	F	41	40	46	72,0	21,6	21,95
79	MO	M	62	129	60	82,8	23,2	4,09
80	MK	F	70	137	44	81,8	22,7	12,68
81	KK	M	53	132	44	94,5	26,7	1,5
82	KMK	F	60	132	44	97,5	28,8	9,78
83	HK	M	55	115	27	87,5	23,4	6,61
84	KN	F	75	221	60	94,5	23,1	17,11
85	NID	F	52	55	43	79,3	22,5	13,09
86	AS	F	71	236	34	89,8	24,4	8,88
87	FS	F	62	130	48	66,0	20,0	20,79
88	MM	M	46	357	13	75,8	20,5	12,45
89	DM	F	52	63	54	74,3	23,2	10,73
90	MI	M	77	76	28	84,0	22,0	1,71

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m ²)	Adiponectina (µg/ml)
91	MKI	F	62	71	60	67,5	19,3	28,04
92	MF	F	69	104	56	66,0	19,1	12,05
93	HT	F	62	128	66	90,5	24,6	16,49
94	MSO	F	41	153	40	89,0	24,9	5,61
95	MT	M	53	237	28	93,0	26,8	5
96	KY	F	64	151	37	91,3	30,2	6,35
97	NYM	M	43	82	42	92,3	23,8	6,24
98	MY	F	63	112	56	67,0	19,4	2,9
99	SK	F	44	49	54	89,8	28,4	16,9
100	OO	M	51	50	48	86,0	23,2	6,91
101	MK	F	71	153	40	82,5	24,0	11,12
102	KT	M	41	895	---	81,8	25,5	9,26
103	KK	F	37	67	59	73,3	22,1	5,4
104	HK	M	41	131	39	96,5	31,9	5,25
105	AYT	M	43	125	29	86,0	25,6	16,5
106	NK	F	60	128	44	70,6	21,0	10,9
107	YN	F	41	79	24	82,5	23,6	4,88
108	MKK	F	44	134	43	73,5	19,1	3,15
109	NMS	F	48	158	27	80,5	24,5	3,03
110	AS	F	63	111	43	83,3	23,8	9,97
111	TK	M	71	133	33	79,0	20,2	3,1
112	RI	M	67	98	29	74,0	19,1	11,25
113	JKT	M	47	217	36	86,3	21,1	5,15
114	MI	M	63	132	40	81,3	21,1	4,46
115	TK	F	57	119	53	90,8	25,4	9,82
116	KAI	F	60	102	59	75,0	19,8	9,55
117	TTM	F	55	119	31	81,5	24,0	2,77
118	YM	M	57	106	59	88,8	22,8	6,68
119	RFY	F	47	66	40	65,5	24	2,4
120	KS	F	50	---	---	81,0	24,1	1,77

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m ²)	Adiponectina (µg/ml)
121	TM	F	60	139	54	117,0	30,9	7,19
122	ES	M	53	153	34	85,3	24,6	8,02
123	PMS	M	41	154	48	75,8	27,8	12,28
124	SM	M	71	284	28	79,8	21,8	2,3
125	MTH	F	45	41	37	75,0	23,6	16,85
126	MM	F	65	116	46	109,8	30,3	3,52
127	TT	M	60	176	42	100,8	31,3	15,65
128	SKT	F	65	434	26	80,5	22,7	4,7
129	TT	M	55	129	35	101,8	30,8	8,75
130	SM	M	42	110	35	92,0	27	6
131	TT	M	46	123	34	86,8	24,6	4,33
132	YM	M	55	214	38	97,0	30,7	8,05
133	SKM	M	52	83	43	82,0	25,1	1,55
134	OM	M	46	129	36	102,3	33,2	15,14
135	SKT	F	41	110	38	60,0	17,8	25,17
136	SH	F	61	120	49	59,0	17,8	6,91
137	TI	M	45	103	37	82,0	23,3	7,12
138	TK	F	69	139	39	90,0	22,9	3,17
139	MK	M	76	59	57	76,4	18,6	27,52
140	TSK	M	40	71	29	87,3	25,1	13,25
141	YM	F	74	159	50	91,0	27,5	16,49
142	JS	M	41	137	29	89,0	23,2	1,4
143	SEH	F	45	68	56	83,3	25,2	9,03
144	NTN	F	46	147	51	96,3	29,5	5,95
145	LS	F	43	71	48	73,5	20,2	25,14
146	MSY	M	56	49	54	71,5	17,4	11,33
147	FM	F	50	307	37	90,8	27,4	1,8
148	IM	M	57	67	41	86,8	25,3	3,36
149	YNM	F	61	215	49	87,5	22,0	8,91
150	SY	F	73	144	43	91,5	27,5	4,65

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m²)	Adiponectina (µg/ml)
151	YM	F	76	85	53	83,0	21,6	11,1
152	NYM	F	44	51	46	75,3	25,2	6,3
153	TT	F	50	74	74	73,3	21,1	12,05
154	TK	F	84	93	39	79,0	23,3	7,87
155	TK	M	41	140	37	94,5	27,4	9,33
156	TN	F	67	314	38	81,0	25,6	6,65
157	LT	F	60	132	48	81,5	22,7	11,2
158	HN	F	68	138	49	91,0	29,1	18,88
159	CK	M	61	104	33	83,5	22,5	4,15
160	TSK	F	62	123	63	80,8	24,0	13,4
161	EH	M	44	178	40	85,0	22,0	2,4
162	YT	F	45	162	52	88,0	23,0	4,25
163	JHKC	F	41	167	41	83,0	25,0	6,59
164	AM	F	67	204	35	98,0	29,2	6
165	SK	M	64	58	65	74,0	20,2	9,87
166	KK	F	69	127	46	80,8	24,9	9,79
167	KT	M	60	155	40	96,5	27,6	12,76
168	ST	M	44	131	51	99,8	30,2	5,25
169	YO	F	64	181	33	94,5	26,2	3,19
170	RFY	M	68	136	44	89,0	24,0	7,25
171	KMS	F	40	66	65	74,8	23,8	6,85
172	TM	M	46	64	38	82,8	25,3	11,67
173	ST	F	64	145	41	100,8	29,2	4,4
174	TN	M	65	215	39	88,8	24,2	3,05
175	SEH	F	57	268	35	88,5	27,7	7,91
176	TS	M	50	154	33	84,5	24,8	5,35
177	TTH	F	40	76	39	76,8	22,9	6,33
178	OO	M	47	206	30	81,5	22,3	4,42
179	MAO	F	56	128	39	74,5	22,9	6,65
180	JTT	M	59	166	19	93,5	27,1	3,35

Número	Iniciais	Sexo	Idade (anos)	Triglicérides (mg/dl)	HDL (mg/dl)	Cintura (cm)	IMC (kg/m²)	Adiponectina (µg/ml)
181	SKT	F	76	88	58	75,5	19,3	11,8
182	PZ	M	44	178	41	79,8	25,2	3,99
183	KM	M	58	209	30	89,3	29,4	2,51
184	TK	M	47	176	38	92,5	26,0	4,35
185	OSIK	F	43	182	38	85,3	27,8	6,7
186	FMA	F	55	106	50	89,3	26,0	4,85
187	ST	M	67	90	33	74,8	19,3	14,8
188	TT	F	60	92	31	84,0	27,2	3,35
189	TS	M	73	355	41	85,8	22,4	6,65
190	YN	F	76	139	61	77,3	19,5	20
191	THT	F	74	255	36	66,3	19,1	9,28
192	NT	F	75	81	48	87,8	25,7	13
193	MKM	F	54	77	53	81,8	23,5	11,38
194	MAI	F	46	124	41	93,1	29,8	9,73
195	MMT	F	56	92	35	95,3	27,9	12,74
196	YUS	F	46	128	59	72,0	21,8	37,5
197	MN	M	50	83	50	88,3	25,8	4,28
198	KK	F	61	122	46	74,8	19,9	4,68
199	HSU	F	47	170	51	93,5	31,7	24,35
200	YNU	F	70	157	42	102,5	32,4	9,45
201	YM	F	63	277	60	91,0	30,6	1,9
202	MM	M	45	985	-	88,0	26,7	3,83
203	KN	M	64	112	47	82,8	22,9	4,51
204	MK	M	70	161	34	90,8	24,7	4,08
205	TFG	F	41	130	39	89,5	29,6	3,36
206	TFG	F	55	123	36	77,8	23,3	9,9
207	HOA	F	59	128	34	87,3	26,3	2,44
208	MM	M	59	222	41	81,8	22,8	14,25
209	KJ	M	52	634	35	95,5	26,0	0,85
210	YO	F	64	94	42	65,0	19,0	13,1

Tabela 2: Características demográficas, antropométricas e tolerância à glicose dos indivíduos ao final do período de seguimento (Artigo 1).

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	Cintura (cm)	Tolerância à glicose
1	TM	M	67	21,6	85,15	TGN
2	HSY	F	71	22,6	94,10	TGD
3	NO	M	69	26,1	98,50	TGN
4	KH	F	68	23,3	83,50	TGN
5	MH	M	74	24,5	85,50	TGN
6	TY	M	52	34,1	101,50	TGD
7	AK	M	61	26,1	85,75	GJA
8	ES	F	68	23,6	89,20	DM
9	ASK	M	58	25,1	88,75	TGD
10	BKK	M	66	23,3	86,25	TGD
11	HI	F	77	22,5	79,00	TGN
12	AAYA	F	56	25,0	83,50	TGD
13	FU	F	62	29,0	96,75	GJA
14	FH	M	65	22,3	82,50	TGD
15	MSY	F	69	22,8	78,50	TGN
16	FO	F	63	24,3	92,50	TGN
17	CHM	M	47	23,4	93,25	GJA
18	HN	F	59	18,8	70,50	GJA
19	AM	F	59	30,0	93,50	DM
20	AST	M	50	28,0	92,00	DM
21	AK	M	53	27,5	91,25	TGD
22	NNM	F	57	24,9	78,50	DM
23	YH	M	62	20,1	72,00	TGD
24	KK	M	56	23,4	85,00	GJA
25	MMM	F	47	24,3	75,50	DM
26	KM	F	76	28,5	93,00	TGD
27	HM	F	63	17,4	70,75	TGN
28	SU	M	66	31,1	101,75	DM
29	KT	M	65	28,2	93,00	TGD
30	HN	F	66	17,3	73,50	GJA

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m²)	Cintura (cm)	Tolerância à glicose
31	EF	M	74	21,8	77,00	GJA
32	YY	M	50	22,7	75,25	TGD
33	SM	M	74	22,7	61,50	TGD
34	JTT	M	56	19,6	79,50	GJA
35	YO	F	66	22,4	79,50	TGN
36	HH	M	54	25,0	92,50	TGD
37	IO	M	67	18,5	75,50	DM
38	HYA	F	49	17,1	53,50	TGN
39	NH	M	66	28,1	93,50	TGN
40	MH	F	68	23,1	81,10	TGN
41	EA	F	64	25,2	80,00	DM
42	SO	M	51	23,0	86,50	TGN
43	JU	M	62	31,5	117,50	GJA
44	NTN	M	68	24,6	84,20	TGD
45	SNM	F	49	26,8	80,50	TGN
46	MM	F	74	18,0	61,00	TGN
47	KK	F	77	21,3	72,50	TGD
48	KOY	F	65	25,8	87,00	TGN
49	JKT	M	52	21,8	72,75	DM
50	HY	F	74	26,9	89,50	DM
51	AI	M	58	28,3	97,50	DM
52	MGK	F	70	29,9	85,50	GJA
53	KY	M	72	20,9	80,00	TGD
54	MK	M	71	20,0	75,00	TGD
55	WK	F	63	19,9	74,00	GJA
56	YS	M	73	25,7	88,50	TGD
57	LYAI	F	56	21,6	66,00	TGN
58	KK	F	75	27,1	83,25	TGN
59	HOA	M	63	21,0	77,50	TGN
60	KK	M	77	28,1	96,00	TGD

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	Cintura (cm)	Tolerância à glicose
61	TT	M	75	23,6	84,50	DM
62	KM	M	61	20,8	72,00	DM
63	KOO	F	53	24,5	75,00	TGD
64	TT	F	69	21,5	68,50	GJA
65	NS	M	61	28,8	100,00	DM
66	IG	M	60	26,9	90,50	DM
67	LU	M	58	25,2	88,00	DM
68	KKT	F	51	21,9	74,50	TGN
69	LKO	M	67	25,8	86,50	TGN
70	IT	M	63	24,4	86,00	GJA
71	HB	F	81	23,3	80,00	TGD
72	KK	M	55	35,4	108,50	DM
73	MI	F	67	26,3	83,00	TGD
74	HH	M	49	25,7	84,00	TGD
75	JO	F	64	32,2	91,00	DM
76	SMK	F	61	19,0	64,50	GJA
77	NTM	M	56	24,3	81,00	TGD
78	NNM	F	47	22,6	72,00	TGN
79	MO	M	66	24,9	86,00	TGD
80	MK	F	76	21,7	77,50	TGD
81	KK	M	59	27,8	90,50	DM
82	KMK	F	66	28,1	90,00	DM
83	HK	M	61	24,1	83,00	DM
84	KN	F	82	22,9	85,00	GJA
85	NID	F	59	24,6	74,50	TGN
86	AS	F	77	25,8	82,00	DM
87	FS	F	68	20,4	64,00	TGN
88	MM	M	52	20,1	73,00	GJA
89	DM	F	58	23,3	70,50	TGN
90	MI	M	82	21,6	79,00	GJA

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m²)	Cintura (cm)	Tolerância à glicose
91	MKI	F	68	18,5	65,00	TGN
92	MF	F	75	19,2	67,00	TGN
93	HT	F	68	26,3	82,50	GJA
94	MSO	F	47	24,7	80,00	GJA
95	MT	M	59	26,8	97,00	TGD
96	KY	F	70	29,5	94,00	TGD
97	NYM	M	49	22,6	84,00	GJA
98	MY	F	69	22,0	71,50	TGD
99	SK	F	50	31,8	89,00	DM
100	OO	M	58	23,2	84,00	DM
101	MK	F	77	23,5	73,50	TGN
102	KT	M	47	23,3	81,00	TGN
103	KK	F	43	21,9	66,00	TGD
104	HK	M	46	38,4	108,00	DM
105	AYT	M	49	26,7	85,50	TGD
106	NK	F	66	19,7	66,50	TGD
107	YN	F	47	25,7	82,00	DM
108	MKK	F	50	19,1	65,00	GJA
109	NMS	F	55	24,5	77,00	DM
110	AS	F	69	24,6	77,50	DM
111	TK	M	76	20,1	74,50	GJA
112	RI	M	74	18,4	71,00	TGD
113	JKT	M	53	21,2	82,00	TGD
114	MI	M	70	21,4	80,00	DM
115	TK	F	64	26,8	85,00	TGD
116	KAI	F	67	20,6	69,00	GJA
117	TTM	F	61	27,2	86,50	DM
118	YM	M	64	23,7	90,00	TGD
119	RFY	F	52	22,8	71,00	TGD
120	KS	F	57	24,0	81,00	DM

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m²)	Cintura (cm)	Tolerância à glicose
121	TM	F	66	31,4	97,00	TGN
122	ES	M	59	24,2	82,00	TGD
123	PMS	M	47	22,7	79,50	TGN
124	SM	M	78	20,0	73,50	GJA
125	MTH	F	51	23,3	73,00	GJA
126	MM	F	72	29,8	97,50	DM
127	TT	M	67	34,5	108,00	TGD
128	SKT	F	71	23,9	79,50	TGD
129	TT	M	61	31,0	104,00	DM
130	SM	M	48	34,6	103,50	GJA
131	TT	M	52	25,8	89,00	DM
132	YM	M	62	30,2	101,00	DM
133	SKM	M	58	23,3	76,50	GJA
134	OM	M	52	33,2	102,00	DM
135	SKT	F	50	17,9	60,00	TGN
136	SH	F	67	18,8	64,50	TGN
137	TI	M	51	23,4	77,50	TGD
138	TK	F	75	24,1	76,00	TGN
139	MK	M	83	18,7	72,00	TGD
140	TSK	M	46	29,0	99,00	TGD
141	YM	F	80	26,4	85,00	TGD
142	JS	M	49	23,2	81,00	TGD
143	SEH	F	52	26,1	80,50	TGD
144	NTN	F	53	29,4	92,00	DM
145	LS	F	49	18,8	66,50	TGD
146	MSY	M	62	16,7	70,00	GJA
147	FM	F	56	28,2	85,00	TGD
148	IM	M	63	25,8	86,00	TGD
149	YNM	F	66	20,4	71,00	TGN
150	SY	F	79	22,4	83,00	TGD

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m²)	Cintura (cm)	Tolerância à glicose
151	YM	F	82	24,6	84,00	TGN
152	NYM	F	51	26,9	80,00	TGD
153	TT	F	56	22,1	77,00	TGN
154	TK	F	81	22,0	71,50	TGN
155	TK	M	48	28,6	94,50	DM
156	TN	F	74	25,9	80,00	TGD
157	LT	F	67	23,0	84,00	TGD
158	HN	F	74	27,4	85,00	TGN
159	CK	M	67	22,3	88,00	TGD
160	TSK	F	68	24,6	78,00	DM
161	EH	M	51	22,3	83,50	TGD
162	YT	F	52	23,1	72,00	GJA
163	JHKC	F	47	24,9	75,00	TGN
164	AM	F	74	28,5	89,75	TGD
165	SK	M	71	20,5	70,75	TGN
166	KK	F	75	23,6	74,00	TGN
167	KT	M	66	27,3	97,00	GJA
168	ST	M	50	32,0	103,00	TGD
169	YO	F	71	26,5	83,00	TGN
170	RFY	M	75	21,7	78,00	GJA
171	KMS	F	47	25,9	80,00	TGD
172	TM	M	53	26,2	85,00	TGN
173	ST	F	71	30,4	97,00	TGD
174	TN	M	72	25,5	90,00	TGD
175	SEH	F	53	27,9	86,00	DM
176	TS	M	56	25,0	87,00	GJA
177	TTH	F	46	25,5	81,00	TGN
178	OO	M	53	21,6	79,00	GJA
179	MAO	F	62	23,2	77,00	GJA
180	JTT	M	66	29,1	100,00	TGD

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m²)	Cintura (cm)	Tolerância à glicose
181	SKT	F	83	19,4	69,00	GJA
182	PZ	M	51	26,2	85,00	TGD
183	KM	M	64	26,4	92,00	DM
184	TK	M	53	25,8	91,00	TGD
185	OSIK	F	49	28,5	89,00	TGN
186	FMA	F	61	26,2	84,50	TGD
187	ST	M	73	20,8	85,00	GJA
188	TT	F	66	27,4	88,00	TGD
189	TS	M	80	21,2	81,15	TGD
190	YN	F	83	20,9	83,00	TGN
191	THT	F	81	19,5	76,00	GJA
192	NT	F	82	23,6	78,00	GJA
193	MKM	F	60	24,0	77,00	TGN
194	MAI	F	52	29,6	94,00	GJA
195	MMT	F	63	29,2	96,00	DM
196	YUS	F	53	22,3	74,00	GJA
197	MN	M	57	24,3	81,00	DM
198	KK	F	68	18,0	71,00	GJA
199	HSU	F	53	34,0	98,00	TGD
200	YNU	F	77	33,1	97,00	TGD
201	YM	F	70	32,2	95,00	TGD
202	MM	M	51	27,8	95,00	TGN
203	KN	M	71	23,6	83,00	TGD
204	MK	M	77	23,5	87,50	TGD
205	TFG	F	48	23,9	75,00	TGN
206	TFG	F	49	20,8	70,00	TGN
207	HOA	F	62	27,0	86,00	TGD
208	MM	M	66	21,3	79,00	TGD
209	KJ	M	59	26,7	100,50	GJA
210	YO	F	71	18,2	58,00	TGN

Tabela 3: Características demográficas, antropométricas, HOMA-R, níveis de adiponectina plasmática e genótipo dos indivíduos portadores de DM (Artigo 2).

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
1	AG	M	77	26,1	2,49	7,45	G/G	G/G	A/G	----	85,50
2	AI	M	66	24,6	1,81	22,50	T/G	G/T	A/G	----	87,50
3	AI	M	58	28,3	2,23	1,00	T/T	G/G	A/A	----	97,50
4	AIK	M	35	33,2	8,94	3,45	T/G	G/G	A/G	----	104,00
5	AM	F	59	30,1	2,87	6,00	T/T	G/T	A/A	----	93,50
6	AMI	F	44	22,9	1,78	3,15	T/G	G/G	A/G	----	78,00
7	AS	F	65	22,6	2,49	12,30	T/T	T/T	A/A	----	69,00
8	AS	M	42	38,0	7,99	5,30	T/T	G/G	A/G	----	118,50
9	AST	M	50	28,0	4,74	7,20	T/T	G/T	A/A	----	92,00
10	ATI	M	50	35,2	10,27	3,95	T/G	G/T	A/G	----	109,00
11	ATN	M	55	26,3	3,67	4,85	T/T	G/G	A/G	----	88,50
12	BM	F	71	22,7	2,36	9,95	T/T	G/T	A/A	----	89,00
13	CAK	F	65	24,5	1,82	1,40	T/G	G/G	A/G	----	82,00
14	CEY	F	65	23,4	4,44	5,50	T/T	G/G	A/A	----	76,00
15	CHN	F	66	24,8	9,62	7,95	T/G	G/T	A/G	----	78,50
16	CMK	M	56	25,6	6,27	3,25	T/T	G/T	A/A	----	86,00
17	CU	M	58	27,1	2,01	1,70	T/G	G/G	A/G	----	91,00
18	EA	F	72	25,1	1,09	5,95	G/G	G/G	G/G	----	95,10
19	EA	F	64	25,2	4,05	4,50	T/G	G/G	A/G	----	80,00
20	EEA	F	47	27,0	5,06	3,90	T/T	G/T	A/A	----	87,50
21	EK	F	63	29,9	10,93	7,20	T/T	G/G	A/A	----	89,50
22	EK	F	55	24,6	2,90	12,50	T/T	T/T	A/A	----	87,50
23	EKS	F	68	29,6	17,46	9,85	T/G	G/G	A/A	----	103,00
24	EMFA	F	36	31,8	3,27	2,55	T/T	G/G	A/A	----	89,00
25	ENI	F	51	26,9	3,14	10,55	T/G	G/G	A/G	----	87,00
26	ES	M	50	30,7	7,96	12,55	T/G	G/G	A/G	H241P	100,50
27	ESHO	F	43	32,5	4,95	11,10	T/T	G/G	A/A	----	90,00
28	ESS	M	42	26,0	3,53	0,90	T/G	G/G	A/A	----	89,00
29	EYY	F	52	26,1	3,14	6,20	T/G	G/T	A/A	----	90,00
30	FFW	F	68	22,8	4,26	8,75	T/G	G/T	A/A	----	81,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
31	FNM	M	55	29,0	5,70	1,85	T/G	G/G	A/A	----	95,50
32	FTK	M	41	----	4,14	1,90	G/G	G/G	G/G	----	----
33	FY	M	56	26,8	13,35	8,40	T/G	G/T	A/A	----	90,00
34	GMUM	F	49	24,0	2,67	0,95	T/T	G/G	A/A	----	77,00
35	GT	M	57	24,8	4,20	4,90	T/G	G/G	A/G	----	91,00
36	HF	M	39	38,8	4,73	3,10	T/G	G/T	A/A	----	119,00
37	HH	M	50	27,5	3,56	2,70	T/T	G/T	A/A	R221S	89,00
38	HK	M	61	24,2	3,55	4,25	T/G	G/G	A/G	----	83,00
39	HN	F	66	25,3	2,93	8,50	T/T	G/G	A/A	----	84,75
40	HO	M	48	30,8	5,05	5,05	T/G	G/T	A/A	----	101,00
41	HU	F	62	30,5	6,62	3,00	T/G	G/G	A/A	----	99,00
42	HY	F	74	27,0	3,96	5,65	T/G	G/G	----	----	89,50
43	HY	F	55	25,7	4,57	1,45	T/G	G/G	A/A	----	83,75
44	HYH	M	40	27,5	5,42	2,55	T/T	G/T	A/A	----	87,00
45	IG	M	59	26,9	4,34	1,75	T/T	G/G	A/A	----	90,50
46	IH	M	73	24,6	2,78	6,30	T/T	G/T	A/A	----	83,50
47	IK	M	53	30,1	6,29	2,50	T/T	G/G	A/A	----	90,50
48	IO	M	71	23,0	2,63	15,50	T/T	G/G	A/A	----	79,50
49	ISS	F	47	24,7	3,70	19,10	T/G	G/T	A/A	----	75,00
50	JAT	M	47	32,8	3,43	3,55	T/T	G/T	A/A	----	104,00
51	JCN	M	66	26,8	4,59	4,00	T/G	G/G	A/G	----	89,00
52	JJK	M	47	29,7	3,66	8,20	T/G	G/T	A/G	----	94,50
53	JN	M	71	27,7	4,04	8,50	T/G	G/G	A/G	----	98,00
54	JN	M	59	28,5	5,58	4,30	G/G	G/G	A/G	----	99,25
55	JO	F	64	32,2	3,44	11,10	T/T	G/T	A/A	----	91,00
56	JOS	F	39	36,5	5,62	2,25	T/G	G/T	A/A	----	101,00
57	KH	M	54	27,0	6,75	3,50	T/T	G/T	A/A	----	86,50
58	KH	F	65	26,0	12,11	6,55	T/G	G/G	----	----	82,00
59	KHN	F	64	20,4	2,63	6,35	T/T	G/G	A/A	----	71,00
60	KK	M	41	34,8	4,17	8,20	T/G	G/T	A/A	----	116,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
61	KK	M	75	28,9	2,13	2,65	T/T	T/T	A/A	----	95,50
62	KK	M	75	24,1	3,08	11,50	T/T	G/T	A/A	----	90,00
63	KK	M	59	27,8	3,17	1,50	T/T	G/G	A/A	----	90,50
64	KK	M	51	35,4	22,47	1,85	G/G	G/G	G/G	----	108,50
65	KK	M	57	26,7	2,96	3,30	T/T	G/T	A/A	----	87,50
66	KMK	F	66	28,2	3,77	3,40	T/T	G/T	A/A	----	90,00
67	KN	M	60	32,8	4,98	6,60	T/T	G/T	A/A	----	103,00
68	KSYS	F	43	31,1	6,46	7,00	T/G	G/G	A/G	----	88,00
69	KT	M	45	33,3	22,04	1,45	T/G	G/T	A/A	----	111,00
70	KT	M	47	26,8	9,80	5,60	T/T	G/T	A/A	----	96,00
71	KU	M	53	28,6	16,71	1,90	T/G	G/T	A/A	----	97,00
72	KY	F	61	21,5	3,89	7,25	T/G	G/G	A/A	----	75,50
73	LH	F	50	26,5	3,73	1,10	T/T	G/T	A/A	R221S	80,00
74	LKM	F	49	29,9	6,25	2,40	T/G	G/T	A/A	----	91,00
75	LSS	F	64	25,5	8,40	6,00	T/G	G/T	A/A	----	83,00
76	LTC	F	50	25,8	2,71	3,20	T/G	G/T	A/A	----	85,00
77	LU	M	58	25,2	5,71	5,85	T/T	T/T	A/A	----	88,00
78	LY	F	52	30,3	5,82	13,95	T/G	G/G	A/A	----	91,00
79	LYK	M	65	25,8	3,81	2,20	T/T	T/T	A/A	----	85,00
80	MA	M	35	28,7	3,64	3,75	T/T	G/T	A/A	----	89,00
81	MA	M	37	25,8	2,31	9,00	G/G	G/G	G/G	----	90,50
82	MAY	F	33	29,3	5,19	1,10	T/T	G/T	A/A	----	77,50
83	MFM	F	31	22,2	14,56	18,00	T/G	G/T	A/G	----	70,00
84	MFM	F	36	27,4	2,92	5,70	T/G	G/G	A/G	----	84,00
85	MH	M	55	31,1	6,26	17,00	T/T	G/T	A/A	----	104,50
86	MHG	F	48	31,2	2,80	10,00	T/T	G/T	A/A	----	97,00
87	MHS	F	69	25,8	2,97	3,60	T/G	G/T	A/G	----	95,00
88	MHS	M	56	26,3	3,05	2,65	T/G	G/G	A/G	----	91,00
89	MJYJ	F	43	27,8	7,00	3,65	T/G	G/G	A/G	----	84,00
90	MK	F	59	25,0	2,75	4,45	T/T	G/G	A/A	----	74,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
91	MK	M	68	27,9	2,31	3,75	T/G	G/T	A/A	----	100,00
92	MK	M	64	25,8	3,18	5,15	T/T	G/G	A/A	----	77,50
93	MK	M	49	25,6	3,18	30,40	T/T	G/G	A/A	----	87,50
94	MK	F	69	24,6	4,50	12,35	T/T	G/G	A/A	----	90,00
95	MKBNS	F	51	27,9	6,23	6,20	T/G	G/T	----	----	88,00
96	MKG	F	60	23,2	3,05	5,30	T/G	G/T	A/G	----	76,50
97	MKK	M	45	26,2	2,63	16,65	T/G	G/T	----	----	93,00
98	MKT	M	35	31,1	8,81	2,20	T/G	G/T	A/G	----	103,00
99	MLFM	F	42	24,8	3,90	4,55	T/T	G/G	A/A	----	75,00
100	MM	F	72	29,8	3,00	9,80	T/T	T/T	A/A	----	97,50
101	MM	F	50	27,7	4,94	4,60	T/T	T/T	A/A	----	82,00
102	MMM	F	47	24,3	2,91	5,45	T/T	G/T	A/A	----	75,50
103	MMN	M	41	23,1	2,67	9,35	T/G	G/G	A/G	H241P	86,00
104	MNC	F	58	25,5	3,91	2,85	T/T	G/T	A/A	----	82,00
105	MNN	F	74	23,4	6,88	5,50	T/T	G/G	A/A	----	78,00
106	MO	F	48	27,5	5,54	7,70	T/G	G/T	A/G	----	97,00
107	MS	F	56	26,0	4,61	11,00	T/G	G/T	A/G	----	84,00
108	MT	M	46	28,2	4,32	5,55	T/G	G/T	A/G	----	89,50
109	MTJT	F	47	23,2	2,52	4,35	T/T	G/T	----	I164T	80,00
110	MTS	F	46	29,9	3,35	4,00	T/T	G/T	----	----	92,00
111	MTT	F	52	28,0	9,31	1,10	T/T	G/T	A/A	----	91,00
112	MTY	M	39	28,9	2,58	1,60	T/T	G/G	A/A	----	91,50
113	MY	F	54	27,2	10,91	6,30	T/T	G/T	A/A	----	89,00
114	MY	M	49	29,8	5,09	1,95	T/T	G/G	A/A	----	98,00
115	MY	F	55	25,9	6,14	9,20	T/G	G/G	A/G	----	80,00
116	MY	F	40	20,9	4,77	7,10	T/T	G/G	A/A	----	74,00
117	MYH	M	49	30,9	2,00	1,95	T/G	G/T	----	----	98,00
118	NF	F	67	22,8	2,07	6,70	T/T	G/G	A/A	----	84,00
119	NK	F	60	31,4	3,20	5,95	T/G	G/T	A/G	----	97,50
120	NKD	F	59	24,2	3,01	9,60	T/G	G/T	A/A	----	80,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
121	NM	F	57	26,4	3,40	12,50	T/T	G/G	A/A	----	81,00
122	NMS	F	55	24,5	2,63	8,70	T/T	G/T	A/A	----	77,00
123	NN	F	57	25,0	2,13	29,00	T/T	G/G	A/A	----	78,50
124	NSJ	M	53	25,4	3,61	1,60	T/G	G/T	A/A	----	95,50
125	NTK	M	55	29,4	1,45	2,60	T/G	G/G	A/G	----	100,00
126	NTN	F	53	29,4	4,11	5,96	T/G	G/T	A/A	----	92,00
127	OD	M	61	24,7	4,99	2,90	T/G	G/G	A/G	----	87,00
128	OM	M	52	33,2	3,03	9,20	T/G	G/T	A/G	----	102,00
129	OO	F	58	23,3	2,58	8,70	T/G	G/T	A/G	----	84,00
130	PTI	M	47	28,3	11,87	4,20	T/T	G/T	A/A	----	95,00
131	QIK	M	53	30,3	8,06	1,45	G/G	G/G	G/G	----	97,00
132	QY	F	47	34,4	3,41	11,55	T/T	G/T	A/G	----	88,00
133	RKK	M	32	28,1	2,65	1,45	T/T	G/G	A/A	----	90,00
134	RMF	M	38	27,6	2,07	1,25	T/T	G/G	----	----	91,00
135	RSG	F	32	22,4	2,38	7,90	T/G	G/G	A/G	H241P	76,00
136	RY	F	70	24,4	2,01	9,15	T/T	G/G	A/A	----	76,50
137	SA	M	50	27,8	5,29	2,60	T/G	G/G	A/G	----	100,00
138	SA	F	53	28,0	11,46	7,91	T/G	G/T	A/G	----	86,00
139	SG	F	72	20,8	3,84	6,65	T/T	G/G	A/A	----	69,00
140	SH	M	52	29,7	6,06	14,00	T/G	G/G	A/A	----	104,00
141	SH	F	68	32,6	6,72	10,95	T/T	G/T	A/A	----	91,00
142	SH	F	66	32,8	2,85	11,80	T/G	G/T	A/G	----	94,50
143	SH	M	53	25,6	3,53	2,75	T/T	G/T	A/A	----	90,00
144	SHY	M	41	40,8	10,19	3,40	T/G	G/T	A/G	----	129,00
145	SI	M	52	23,7	5,19	3,10	T/G	G/T	A/G	I164T	87,00
146	SK	F	60	25,4	9,30	4,40	G/G	G/G	G/G	----	90,00
147	SK	M	63	22,5	2,47	3,00	T/G	G/T	A/G	----	86,00
148	SK	M	66	26,8	9,55	2,50	T/G	G/G	A/G	----	96,00
149	SK	F	50	31,9	2,62	25,70	T/T	T/T	A/A	----	89,00
150	SM	F	65	25,5	6,05	5,10	T/G	G/T	A/A	----	85,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
151	SM	F	69	25,1	2,26	14,40	T/G	G/T	A/G	----	82,00
152	SM	M	66	27,2	4,29	3,90	T/T	G/G	A/G	----	100,00
153	SNA	F	44	24,0	2,28	2,60	T/G	G/G	A/G	----	78,00
154	SNF	F	51	28,2	9,14	3,25	T/T	G/T	A/A	----	91,00
155	SO	M	54	21,8	2,91	5,75	T/G	G/G	A/G	----	80,00
156	SO	M	73	28,7	2,68	5,50	T/T	G/T	A/A	----	94,50
157	SS	F	65	26,4	2,86	10,65	T/G	G/T	A/G	----	92,50
158	ST	M	54	28,9	7,90	6,70	G/G	G/G	G/G	----	100,00
159	SY	M	48	24,4	2,15	2,70	T/T	G/T	A/A	----	82,00
160	TF	M	61	24,9	5,99	5,30	T/G	G/G	A/G	----	91,50
161	TG	M	45	26,2	3,83	6,70	T/G	G/T	A/A	----	96,00
162	TGK	F	52	27,8	5,84	5,40	T/T	G/T	A/A	----	88,20
163	TH	M	59	34,1	7,97	4,30	T/T	T/T	A/A	----	108,00
164	TIM	F	47	26,7	2,43	9,55	T/G	G/T	A/G	----	80,00
165	TJCI	M	45	21,5	3,58	3,80	T/G	G/T	A/A	----	85,00
166	TK	M	61	27,3	3,11	8,70	T/T	G/T	----	----	96,25
167	TM	M	62	26,6	3,29	1,65	T/G	G/G	A/A	----	98,00
168	TN	F	44	35,8	10,97	2,40	T/G	G/T	A/A	----	95,00
169	TO	M	66	25,5	2,87	4,45	T/T	G/T	A/A	----	89,00
170	TOF	F	76	33,7	2,33	17,00	T/T	T/T	A/A	----	104,00
171	TS	M	59	23,4	2,22	4,60	T/T	G/T	A/A	----	82,00
172	TSY	F	51	31,1	5,37	9,75	T/G	G/G	A/G	----	96,00
173	TT	M	76	26,4	2,09	5,40	T/G	G/T	A/G	----	94,00
174	TT	M	74	32,5	3,22	9,90	T/T	G/G	A/A	----	105,00
175	TT	M	46	27,4	2,97	6,15	T/T	G/T	A/A	----	87,00
176	TT	M	75	23,7	4,19	5,27	T/G	G/G	A/A	----	84,50
177	TTI	F	68	37,9	3,76	8,00	T/T	G/T	A/A	----	95,00
178	TTM	F	61	27,2	5,15	2,77	T/G	G/T	A/G	----	86,50
179	TTMM	F	48	29,7	2,97	3,10	T/T	G/G	A/A	----	87,00
180	TUI	F	70	21,8	2,69	8,55	T/T	G/T	A/A	----	75,50

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
181	TUS	F	53	25,6	2,98	8,15	G/G	G/T	A/A	----	84,00
182	TYSJ	F	45	30,0	7,84	9,90	T/G	G/T	A/G	----	94,00
183	VM	M	39	32,0	5,21	3,40	T/G	G/T	A/G	----	104,00
184	VOF	F	70	24,7	5,24	10,50	T/G	G/T	A/A	----	87,00
185	WTM	M	45	27,6	4,89	3,50	T/G	G/T	A/A	----	96,00
186	YA	M	59	23,5	2,26	3,60	T/G	G/G	A/G	----	88,00
187	YK	M	62	26,6	2,87	3,90	T/T	G/T	A/G	----	94,00
188	YK	M	50	21,5	2,02	15,50	T/T	G/T	A/G	----	84,00
189	YK	F	68	24,4	2,54	5,45	T/T	G/T	A/G	----	88,00
190	YM	M	62	30,2	2,26	5,25	T/T	G/G	A/A	----	101,00
191	YMO	F	62	22,1	15,90	11,00	G/G	G/G	G/G	----	80,00
192	YN	M	61	33,6	2,65	3,25	T/G	G/T	A/G	----	104,00
193	YS	F	58	27,9	11,40	6,45	G/G	G/G	G/G	----	86,50
194	YS	M	73	32,0	2,07	27,50	T/T	G/T	A/A	----	102,50
195	YT	F	69	26,0	2,91	6,00	T/G	G/G	A/A	----	85,00
196	YT	M	56	32,1	8,30	2,25	T/G	G/T	A/A	----	107,00
197	YT	F	62	25,5	2,68	4,50	T/T	T/T	A/G	----	76,00
198	YUM	F	35	23,1	2,39	3,00	T/T	G/T	A/A	----	90,00
199	YY	M	61	24,5	2,68	3,45	T/G	G/T	A/A	----	92,50
200	ZH	M	69	22,2	3,06	4,75	T/T	G/T	A/A	----	85,00

Tabela 4: Características demográficas, antropométricas, HOMA-R, níveis de adiponectina plasmática e genótipo dos indivíduos com TNG (Atrigo 2).

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
201	AT	F	68	21,69	1,07	6,10	T/T	G/T	A/A	----	73,00
202	AB	F	64	22,91	0,93	11,45	T/G	G/T	A/G	----	78,00
203	AHT	F	48	18,71	0,59	19,80	T/G	G/G	A/G	----	66,50
204	AK	F	47	25,70	1,96	7,30	T/G	G/T	A/G	----	75,50
205	AMHT	F	42	22,43	1,45	9,55	T/T	G/T	A/A	----	74,00
206	AMO	F	47	25,47	0,93	1,70	T/G	G/G	A/G	----	80,50
207	AMY	M	49	20,97	1,11	14,60	T/T	G/T	A/A	----	79,00
208	ANO	F	65	24,29	0,62	12,15	T/T	G/G	A/A	----	80,50
209	AOS	F	64	22,34	1,37	1,90	T/T	G/G	A/A	----	80,00
210	ASY	M	32	21,00	1,41	2,25	T/G	G/T	A/G	----	78,00
211	AT	M	62	23,08	0,80	12,85	T/G	G/G	A/G	----	80,50
212	ATA	M	48	22,00	0,70	3,15	G/G	G/G	G/G	H241P	80,50
213	ATFK	F	40	15,89	1,46	12,00	G/G	G/G	G/G	----	62,00
214	AY	F	43	27,66	1,74	8,35	T/G	G/G	A/G	----	87,00
215	AYS	M	41	22,41	1,03	3,10	T/G	G/T	A/G	----	78,50
216	CM	F	30	20,92	1,28	20,00	T/T	G/T	A/A	----	73,00
217	CNS	M	49	19,42	0,97	13,20	T/G	G/T	A/G	----	77,00
218	CSK	M	42	18,14	1,62	19,00	T/T	G/T	A/A	----	74,50
219	CTTI	F	47	23,93	0,83	5,25	T/G	G/T	A/G	----	77,50
220	CU	M	45	30,11	1,61	2,35	T/T	G/G	A/A	----	97,00
221	DM	F	58	23,37	1,96	8,95	T/T	G/G	A/A	----	70,50
222	EEK	F	39	18,41	0,52	3,45	G/G	G/G	G/G	----	62,00
223	EHHT	F	41	19,48	1,99	4,60	T/G	G/T	A/G	----	79,50
224	EHI	M	42	22,46	0,39	11,40	G/G	G/G	A/G	----	80,00
225	EIG	M	51	25,29	1,72	7,00	T/G	G/G	A/G	----	80,00
226	EMG	M	34	27,18	1,42	3,40	T/T	G/G	A/A	----	85,00
227	EMI	F	56	24,39	1,86	7,00	T/T	G/T	A/A	----	81,00
228	ET	F	58	26,18	0,97	16,35	T/G	G/T	A/G	----	78,00
229	ETU	F	46	28,15	1,14	4,35	T/G	G/G	A/G	----	83,00
230	EU	F	40	20,22	0,67	4,50	T/T	G/G	A/A	----	78,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
231	EY	F	51	23,90	1,04	10,90	T/G	G/T	A/G	----	80,00
232	FMOO	F	42	21,97	0,25	3,65	T/G	G/T	A/G	----	70,00
233	FNG	F	37	20,26	0,41	19,50	T/G	G/T	A/G	----	76,00
234	FO	F	55	20,39	0,86	5,75	T/T	G/T	A/A	----	73,00
235	FO	F	63	24,35	----	5,60	T/T	G/G	A/A	----	92,50
236	FS	F	68	20,47	1,54	10,50	T/G	G/G	A/G	----	64,00
237	FS	M	62	26,53	1,58	1,00	G/G	G/G	G/G	----	92,00
238	FU	F	55	22,43	3,35	10,40	T/T	G/G	A/G	----	70,75
239	HÁ	M	53	23,40	1,24	1,85	T/T	G/G	A/A	----	81,00
240	HE	M	70	23,03	0,43	6,00	T/G	G/T	A/A	----	87,00
241	HI	F	77	22,55	1,41	8,40	T/G	G/G	A/G	----	79,00
242	HI	F	54	24,19	1,16	13,70	T/T	G/T	A/A	----	84,50
243	HK	M	70	19,71	0,83	3,20	T/T	G/T	A/A	----	78,00
244	HKO	F	50	25,25	1,68	4,25	T/T	G/G	A/A	----	84,50
245	HKTO	F	42	20,55	1,05	24,35	T/T	G/G	A/A	----	65,00
246	HM	F	63	17,46	----	20,00	G/G	G/G	G/G	----	70,75
247	HMB	F	52	23,20	1,29	3,25	T/T	G/T	A/A	----	76,50
248	HMI	M	44	23,62	1,65	6,15	T/G	G/G	A/G	----	89,10
249	HMK	F	49	23,85	1,69	6,65	G/G	G/G	G/G	----	81,00
250	HN	F	74	27,40	1,81	21,70	T/T	G/T	A/A	----	85,00
251	HO	M	72	23,30	1,48	5,85	T/G	G/G	A/G	----	86,00
252	HO	M	60	22,96	0,85	7,15	T/G	G/G	A/G	----	81,00
253	HO	M	63	21,02	1,75	5,60	T/G	G/G	A/G	----	77,50
254	HS	F	45	21,41	0,59	6,30	T/G	G/G	A/G	----	74,00
255	HTG	F	59	24,25	----	2,10	T/T	G/G	A/A	R221S	80,50
256	IA	F	59	21,66	0,27	13,95	T/T	G/T	A/A	----	71,00
257	IMAF	F	45	22,43	1,60	10,30	T/T	G/G	A/A	----	71,00
258	IST	F	63	17,98	0,69	4,45	T/G	G/G	A/G	----	74,50
259	IT	M	40	27,91	0,61	1,10	T/G	G/T	A/G	----	87,00
260	IYST	F	33	23,65	1,24	19,75	T/T	G/T	A/A	----	68,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
261	JHKC	F	47	24,98	1,55	14,75	T/T	G/T	A/A	----	75,00
262	JKK	F	42	24,50	0,66	13,35	T/G	G/T	A/G	----	78,00
263	JKN	M	45	25,55	2,45	2,25	T/T	G/G	A/A	----	92,00
264	JMNS	F	44	22,80	1,83	8,80	T/T	G/G	A/A	----	70,00
265	JTK	M	44	25,47	0,83	9,30	T/G	G/G	A/A	----	80,75
266	JTT	M	41	27,02	1,32	9,28	T/G	G/G	A/G	----	89,00
267	KH	F	70	18,60	0,76	21,25	T/G	G/G	----	----	63,00
268	KH	F	68	23,32	1,38	13,00	T/G	G/T	A/A	----	83,50
269	KHK	F	54	27,53	1,82	27,85	T/T	G/T	----	----	77,50
270	KK	F	75	27,06	1,28	10,00	G/G	G/G	A/A	----	83,25
271	KK	F	79	20,41	0,43	12,50	T/T	G/T	A/A	----	73,50
272	KK	F	86	21,84	1,19	24,15	T/T	G/T	A/A	----	78,00
273	KKT	F	51	21,98	1,69	10,80	T/G	G/T	A/G	----	74,50
274	KM	M	46	20,55	1,85	7,35	T/T	G/T	A/A	----	81,50
275	KNI	F	50	21,33	0,84	8,50	T/G	G/G	A/G	----	66,00
276	KS	M	80	20,70	0,49	19,50	G/G	G/G	G/G	----	80,00
277	KT	M	47	23,37	0,65	6,90	T/T	G/T	A/A	----	81,00
278	KY	M	67	24,03	0,56	8,85	T/T	G/G	A/A	----	83,00
279	LEM	M	43	22,55	0,63	2,55	T/G	G/T	A/G	----	80,00
280	LKO	M	67	25,87	2,41	5,20	T/T	G/T	A/A	----	86,50
281	LLIK	F	41	24,12	1,67	12,90	T/T	G/G	A/A	----	80,00
282	LMIA	F	53	27,04	1,91	11,90	G/G	G/G	A/G	----	87,50
283	LMOG	F	34	21,68	0,83	11,20	T/G	G/T	A/A	----	77,00
284	LO	M	50	25,36	0,93	5,05	T/T	G/G	A/A	----	82,00
285	LSY	F	44	23,60	1,39	2,50	T/T	G/T	A/A	----	78,00
286	MAN	F	35	25,47	1,33	8,35	T/T	T/T	A/A	----	78,00
287	MF	F	75	19,20	1,31	12,06	T/T	G/G	A/A	----	67,00
288	MH	F	50	21,72	0,74	2,50	T/T	G/T	A/A	----	73,00
289	MH	M	74	24,57	0,43	5,60	T/G	G/G	A/G	----	85,50
290	MK	M	57	24,31	----	1,80	T/T	G/G	A/A	----	82,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
291	MK	M	40	25,22	1,29	5,35	T/G	G/T	A/A	----	85,00
292	MK	F	77	23,56	1,57	8,15	T/G	G/G	A/G	----	73,50
293	MKI	F	68	18,50	0,84	22,30	T/T	G/T	A/A	----	65,00
294	MKM	F	60	24,08	1,98	19,05	T/T	G/T	A/A	----	77,00
295	MLEHT	F	47	25,10	1,07	7,25	G/G	G/G	G/G	----	75,00
296	MM	F	52	23,78	1,03	3,55	T/G	G/T	A/G	----	74,00
297	MM	F	74	18,06	2,12	12,25	T/T	T/T	G/G	----	61,00
298	MM	M	43	21,83	1,10	20,00	T/T	G/G	A/G	----	80,75
299	MMM	F	57	21,53	0,68	25,00	T/G	G/G	A/G	----	71,00
300	MMY	F	44	19,99	0,48	17,00	T/G	G/G	A/G	----	65,00
301	MO	M	64	28,18	1,95	2,15	T/T	G/G	A/A	----	101,75
302	MO	M	66	20,12	0,54	11,25	T/G	G/T	----	----	72,00
303	MOO	F	58	23,31	1,50	17,55	T/G	G/T	A/G	----	75,00
304	MOP	F	70	36,14	----	6,15	T/T	G/T	A/A	----	103,40
305	MS	F	59	20,72	1,20	16,95	T/T	G/T	A/A	----	77,25
306	MSD	F	54	22,77	1,02	5,25	T/G	G/G	A/G	----	73,00
307	MSE	F	48	21,80	1,35	23,65	T/T	G/T	A/A	----	67,25
308	MSM	F	34	21,44	1,03	4,45	T/T	T/T	A/A	----	68,50
309	MSY	F	69	22,88	----	6,55	T/G	G/G	A/G	----	78,50
310	MT	M	32	19,25	1,13	18,40	T/T	G/G	A/A	----	71,00
311	MT	F	54	20,09	0,64	5,85	T/T	G/T	A/A	----	71,00
312	MT	M	66	20,81	1,54	6,30	T/T	G/T	A/A	----	82,00
313	MTI	F	69	25,32	1,20	22,50	T/T	G/G	A/A	----	85,50
314	MTSR	F	41	22,56	1,39	6,90	T/G	G/G	A/G	----	71,50
315	NAM	F	31	18,93	1,41	9,35	T/T	G/T	A/A	----	69,50
316	NF	F	41	20,18	1,56	7,85	T/T	G/T	A/A	----	66,75
317	NF	M	34	21,55	1,50	6,70	T/T	G/T	A/A	----	79,50
318	NH	M	66	28,11	1,20	3,00	T/G	G/T	A/G	----	93,50
319	NID	F	59	24,64	1,59	13,50	T/G	G/T	A/G	----	74,50
320	NK	F	52	22,71	1,66	15,70	T/T	G/T	A/A	----	74,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
321	NM	M	77	24,84	----	4,75	T/T	G/T	A/A	----	87,00
322	NM	M	45	24,61	1,97	2,40	T/T	G/T	A/A	----	88,00
323	NNM	F	47	22,69	0,60	26,80	T/G	G/T	A/G	H241P	72,00
324	NO	F	43	18,76	1,44	15,00	T/G	G/T	A/G	----	64,00
325	NO	M	69	26,15	1,07	7,80	T/T	T/T	A/A	----	98,50
326	NTS	M	36	17,67	0,54	15,75	T/T	G/G	A/A	----	67,00
327	NUS	F	69	26,46	1,35	16,50	T/G	G/G	A/G	----	83,00
328	OAY	M	36	22,84	0,83	6,75	T/T	G/G	A/A	----	81,00
329	OOF	F	62	23,89	1,58	20,65	T/G	G/G	A/G	----	77,00
330	OSIK	F	49	28,51	1,59	9,10	T/T	G/T	A/A	----	89,00
331	PMS	M	47	22,78	1,89	3,25	T/G	G/T	A/G	----	79,50
332	PT	N	49	22,90	1,43	10,00	T/G	G/G	A/G	----	80,00
333	RAM	M	39	23,62	0,77	7,90	T/G	G/G	A/G	----	86,50
334	RFKS	F	36	24,95	1,51	8,00	T/T	G/T	A/A	----	78,00
335	RMHC	F	43	23,85	1,68	5,80	T/T	G/T	A/A	----	70,50
336	RNM	M	33	26,33	1,27	4,75	T/T	G/G	A/A	----	83,50
337	SAUT	F	42	26,32	1,81	7,65	T/G	G/T	G/G	----	79,00
338	SE	F	51	21,94	1,03	18,50	T/T	G/T	A/A	----	75,25
339	SH	F	67	18,80	0,49	6,91	T/T	G/T	----	----	64,50
340	SHK	M	40	21,27	1,13	17,00	T/G	G/T	A/G	----	80,00
341	SIN	F	59	21,78	0,49	17,00	T/T	G/G	A/A	----	71,00
342	SK	F	50	22,68	2,20	6,30	T/G	G/G	A/G	----	77,00
343	SK	M	71	20,52	0,81	19,30	T/T	T/T	A/A	----	70,75
344	SKT	F	50	17,91	0,60	16,00	T/G	G/T	A/G	----	60,00
345	SM	M	39	24,63	2,02	4,65	T/T	G/T	A/A	----	79,00
346	SM	F	64	25,00	1,56	13,50	T/T	G/T	A/A	----	84,00
347	SMMA	F	36	23,99	1,18	21,50	T/G	G/T	A/G	----	77,00
348	SN	F	54	21,57	1,54	6,35	T/G	G/T	A/G	----	70,00
349	SN	M	55	21,98	0,99	8,05	T/T	G/T	A/A	----	74,00
350	SN	M	74	24,34	0,74	20,35	T/G	G/G	A/G	----	94,50

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
351	SO	M	65	19,74	0,80	6,20	T/G	G/G	A/G	----	77,00
352	SS	F	69	19,18	1,57	24,10	T/G	G/G	A/G	----	69,50
353	SSKN	F	35	20,39	1,09	5,45	T/G	G/G	A/G	H241P	64,50
354	SSN	F	80	22,71	1,07	21,00	T/G	G/T	A/G	----	82,25
355	SST	F	54	21,19	1,65	9,90	T/G	G/T	A/G	H241P	74,50
356	ST	F	86	22,05	0,60	12,70	G/G	G/G	G/G	----	87,00
357	SU	F	60	21,64	0,98	21,00	T/T	G/G	A/A	----	79,50
358	SWK	F	60	23,95	1,16	12,80	T/T	G/T	A/A	----	78,00
359	TF	M	64	19,38	1,71	16,00	T/T	G/G	A/A	----	76,00
360	TF	F	49	20,83	0,92	7,25	T/G	G/G	A/G	----	70,00
361	TFG	F	48	23,96	1,04	3,36	T/T	G/G	A/A	----	75,00
362	TH	M	64	18,86	0,25	14,40	T/G	G/G	A/G	----	71,00
363	TH	M	44	23,11	1,43	3,25	T/G	G/G	A/G	----	81,00
364	TH	M	40	26,16	1,75	11,75	T/T	G/T	A/A	----	89,00
365	TI	F	56	22,15	1,74	16,50	T/G	G/T	A/G	----	77,00
366	TK	F	58	18,98	0,89	18,35	T/T	G/T	A/A	----	67,00
367	TK	F	69	21,27	0,72	21,45	G/G	G/G	G/G	----	69,00
368	TK	F	75	24,11	1,89	3,18	T/G	T/G	A/A	----	76,00
369	TK	F	81	22,07	0,92	7,88	T/T	G/G	A/A	----	71,50
370	TKT	F	60	24,30	1,64	5,75	T/T	G/T	A/A	----	86,50
371	TKY	F	47	24,92	0,94	34,00	T/G	G/T	A/G	----	82,00
372	TLNH	F	40	27,86	1,09	5,50	T/G	G/G	A/G	----	80,00
373	TM	M	74	21,18	0,55	9,00	T/T	G/T	A/A	----	76,00
374	TM	M	66	21,16	1,81	26,95	T/T	G/G	----	----	76,00
375	TM	M	67	21,64	1,09	4,30	T/T	G/G	A/A	----	85,15
376	TM	F	54	20,65	1,21	20,00	G/G	G/G	A/G	----	67,00
377	TMKS	F	42	18,97	0,85	15,70	T/G	G/G	A/A	----	72,00
378	TMT	F	34	20,13	0,64	6,80	T/T	T/T	A/A	----	64,00
379	TO	M	59	22,75	1,76	3,00	T/T	G/T	A/A	----	81,00
380	TS	M	85	18,28	0,33	6,70	T/G	G/T	A/G	----	76,00

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	T45G	G276T	A349G	Exon3	Cintura (cm)
381	TT	F	48	19,17	0,22	5,25	T/G	G/G	A/G	----	62,00
382	TTN	F	50	22,60	0,98	6,45	T/G	G/T	A/G	----	80,50
383	TTT	M	36	25,54	0,71	9,00	T/T	G/T	A/A	----	83,00
384	TY	F	85	20,60	0,48	14,80	T/T	G/T	A/A	----	79,50
385	TY	M	63	23,34	0,23	9,45	T/G	G/G	A/G	----	77,00
386	TY	F	70	20,45	1,06	16,65	T/T	G/G	A/A	----	65,50
387	WST	M	46	24,93	0,63	10,40	T/T	G/G	A/A	----	92,00
388	WSY	M	46	24,60	1,84	9,90	T/T	G/G	A/A	----	84,00
389	YA	M	75	24,88	1,44	5,10	T/T	G/T	A/A	----	87,50
390	YF	F	51	25,64	2,09	8,45	T/G	G/G	A/G	----	66,00
391	YI	M	36	22,57	1,45	6,20	T/T	G/T	A/A	----	83,75
392	YI	M	56	23,58	1,28	10,00	G/G	G/G	G/G	----	88,50
393	YLOT	F	39	24,03	1,67	6,20	T/T	G/G	A/A	----	75,00
394	YM	F	82	24,60	1,26	27,50	T/G	G/T	A/G	----	84,00
395	YM	M	59	26,39	0,72	7,55	T/T	G/G	A/A	----	99,00
396	YM	F	48	19,79	1,57	6,95	T/G	G/G	A/G	----	72,00
397	YN	F	83	20,93	1,73	22,80	T/G	G/T	A/G	----	83,00
398	YO	F	71	18,24	0,65	16,00	T/T	T/T	A/A	----	58,00
399	YT	F	58	21,27	1,06	10,50	T/G	G/G	A/G	----	65,00
400	YY	F	55	22,81	1,92	4,90	T/T	G/T	A/A	----	75,00

Tabela 5: Características demográficas, antropométricas e laboratoriais dos indivíduos portadores de DM (em vermelho, os propósitos carreadores da mutação p.Gly63ValfsX106) (Artigo 3).

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
1	AG	M	77	26,1	2,49	7,45	G/G	G/G	A/G
2	AI	M	66	24,6	1,81	22,50	87,50	50	370
3	AI	M	58	28,3	2,23	1,00	97,50	42	680
4	AIK	M	35	33,2	8,94	3,45	104,00	64	825
5	AM	F	59	30,1	2,87	6,00	93,50	52	346
6	AMI	F	44	22,9	1,78	3,15	78,00	51	107
7	AN	M	56	23,9	6,87	2,20	87,75	39	182
8	AS	F	65	22,6	2,49	12,30	69,00	58	182
9	AS	M	42	38,0	7,99	5,30	118,50	56	83
10	AS	M	50	27,8	5,29	2,60	100,00	75	1521
11	AS	F	53	28,0	11,46	7,91	86,00	83	1058
12	AST	M	50	28,0	4,74	7,20	92,00	45	344
13	ATI	M	50	35,2	10,27	3,95	109,00	64	305
14	ATN	M	55	26,3	3,67	4,85	88,50	41	419
15	BM	F	71	22,7	2,36	9,95	89,00	57	325
16	CAK	F	65	24,5	1,82	1,40	82,00	42	301
17	CEY	F	65	23,4	4,44	5,50	76,00	51	166
18	CHN	F	66	24,8	9,62	7,95	78,50	36	349
19	CMK	M	56	25,6	6,27	3,25	86,00	48	76
20	CU	M	58	27,1	2,01	1,70	91,00	44	491
21	EA	F	72	25,1	1,09	5,95	95,10	51	90
22	EA	F	64	25,2	4,05	4,50	80,00	55	238
23	EEA	F	47	27,0	5,06	3,90	87,50	37	287
24	EK	F	63	29,9	10,93	7,20	89,50	39	270
25	EK	F	55	24,6	2,90	12,50	87,50	91	269
26	EKS	F	68	29,6	17,46	9,85	103,00	47	284
27	EMFA	F	36	31,8	3,27	2,55	89,00	43	269
28	ENI	F	51	26,9	3,14	10,55	87,00	54	212
29	ESHO	F	43	32,5	4,95	11,10	90,00	42	177
30	ESS	M	42	26,0	3,53	0,90	89,00	37	130

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
31	EYY	F	52	26,1	3,14	6,20	90,00	54	180
32	FFW	F	68	22,8	4,26	8,75	81,00	43	158
33	FNM	M	55	29,0	5,70	1,85	95,50	40	450
34	FTK	M	41	-	4,14	1,90	-	38	180
35	FY	M	56	26,8	13,35	8,40	90,00	46	120
36	GMUM	F	49	24,0	2,67	0,95	77,00	54	392
37	GT	M	57	24,8	4,20	4,90	91,00	49	328
38	HF	M	39	38,8	4,73	3,10	119,00	43	430
39	HH	M	50	27,5	3,56	2,70	89,00	46	631
40	HK	M	61	24,2	3,55	4,25	83,00	40	117
41	HN	F	66	25,3	2,93	8,50	84,75	47	345
42	HO	F	59	24,2	3,01	9,60	80,00	44	161
43	HU	F	62	30,5	6,62	3,00	99,00	53	157
44	HY	F	74	27,0	3,96	5,65	89,50	49	687
45	HY	M	40	27,5	5,42	2,55	87,00	53	633
46	HYH	M	48	30,8	5,05	5,05	101,00	52	596
47	HYUF	F	55	25,7	4,57	1,45	83,75	42	291
48	IG	M	59	26,9	4,34	1,75	90,50	58	875
49	IH	M	73	24,6	2,78	6,30	83,50	46	267
50	IK	M	53	30,1	6,29	2,50	90,50	44	35
51	IO	M	71	23,0	2,63	15,50	79,50	41	214
52	ISS	F	47	24,7	3,70	19,10	75,00	62	96
53	JAT	M	47	32,8	3,43	3,55	104,00	49	574
54	JCN	M	66	26,8	4,59	4,00	89,00	46	560
55	JJK	M	47	29,7	3,66	8,20	94,50	59	270
56	JN	M	71	27,7	4,04	8,50	98,00	49	215
57	JN	M	59	28,5	5,58	4,30	99,25	47	246
58	JO	F	64	32,2	3,44	11,10	91,00	45	177
59	JOS	F	39	36,5	5,62	2,25	101,00	47	359
60	KH	F	64	20,4	2,63	6,35	71,00	56	167

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
61	KH	M	54	27,0	6,75	3,50	86,50	41	254
62	KH	F	65	26,0	12,11	6,55	82,00	41	324
63	KK	M	41	34,8	4,17	8,20	116,00	41	146
64	KK	M	75	28,9	2,13	2,65	95,50	38	401
65	KK	M	75	24,1	3,08	11,50	90,00	55	172
66	KK	M	59	27,8	3,17	1,50	90,50	51	229
67	KK	M	51	35,4	22,47	1,85	108,50	42	422
68	KK	M	57	26,7	2,96	3,30	87,50	44	211
69	KMK	F	66	28,2	3,77	3,40	90,00	44	171
70	KN	M	60	32,8	4,98	6,60	103,00	43	175
71	KSYS	F	43	31,1	6,46	7,00	88,00	50	315
72	KT	M	45	33,3	22,04	1,45	111,00	34	399
73	KT	M	47	26,8	9,80	5,60	96,00	46	401
74	KU	M	53	28,6	16,71	1,90	97,00	43	345
75	KY	F	61	21,5	3,89	7,25	75,50	101	1815
76	LH	F	50	26,5	3,73	1,10	80,00	53	417
77	LKM	F	49	29,9	6,25	2,40	91,00	52	226
78	LSS	F	64	25,5	8,40	6,00	83,00	51	126
79	LT	F	50	25,8	2,71	3,20	85,00	59	139
80	LU	M	58	25,2	5,71	5,85	88,00	68	1159
81	LY	F	52	30,3	5,82	13,95	91,00	71	131
82	LYK	M	65	25,8	3,81	2,20	85,00	39	310
83	MA	M	35	28,7	3,64	3,75	89,00	38	205
84	MA	M	37	25,8	2,31	9,00	90,50	57	324
85	MAY	F	33	29,3	5,19	1,10	77,50	47	402
86	MF	F	36	27,4	2,92	5,70	84,00	56	206
87	MFM	F	31	22,2	14,56	18,00	70,00	50	137
88	MH	M	55	31,1	6,26	17,00	104,50	53	106
89	MHG	F	48	31,2	2,80	10,00	97,00	63	1067
90	MHS	F	69	25,8	2,97	3,60	95,00	59	409

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
91	MHS	M	56	26,3	3,05	2,65	91,00	44	212
92	MJYJ	F	43	27,8	7,00	3,65	84,00	48	161
93	MK	F	59	25,0	2,75	4,45	74,00	49	119
94	MK	M	68	27,9	2,31	3,75	100,00	40	349
95	MK	M	64	25,8	3,18	5,15	77,50	52	349
96	MK	M	49	25,6	3,18	30,40	87,50	47	210
97	MK	F	69	24,6	4,50	12,35	90,00	32	184
98	MKBNS	F	51	27,9	6,23	6,20	88,00	45	586
99	MKG	F	60	23,2	3,05	5,30	76,50	34	380
100	MKK	M	45	26,2	2,63	16,65	93,00	51	213
101	MKT	M	35	31,1	8,81	2,20	103,00	51	656
102	MLFN	F	42	24,8	3,90	4,55	75,00	47	124
103	MM	F	72	29,8	3,00	9,80	97,50	60	178
104	MM	F	50	27,7	4,94	4,60	82,00	52	213
105	MMM	F	47	24,3	2,91	5,45	75,50	38	257
106	MMN	M	41	23,1	2,67	9,35	86,00	46	226
107	MNC	F	58	25,5	3,91	2,85	82,00	42	256
108	MNN	F	74	23,4	6,88	5,50	78,00	43	134
109	MO	F	48	27,5	5,54	7,70	97,00	47	268
110	MS	F	56	26,0	4,61	11,00	84,00	38	202
111	MT	M	46	28,2	4,32	5,55	89,50	43	252
112	MTJT	F	47	23,2	2,52	4,35	80,00	46	480
113	MTS	F	46	29,9	3,35	4,00	92,00	54	198
114	MTT	F	52	28,0	9,31	1,10	91,00	32	393
115	MTY	M	39	28,9	2,58	1,60	91,50	51	397
116	MY	F	54	27,2	10,91	6,30	89,00	41	233
117	MY	M	49	29,8	5,09	1,95	98,00	47	539
118	MY	F	55	25,9	6,14	9,20	80,00	62	256
119	MY	F	40	20,9	4,77	7,10	74,00	43	69
120	MYH	M	49	30,9	2,00	1,95	98,00	46	218

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
121	NF	F	67	22,8	2,07	6,70	84,00	59	434
122	NK	F	60	31,4	3,20	5,95	97,50	34	270
123	NM	F	57	26,4	3,40	12,50	81,00	44	308
124	NMS	F	55	24,5	2,63	8,70	77,00	41	197
125	NN	F	57	25,0	2,13	29,00	78,50	56	58
126	NSJ	M	53	25,4	3,61	1,60	95,50	38	399
127	NTK	M	55	29,4	1,45	2,60	100,00	37	171
128	NTN	F	53	29,4	4,11	5,96	92,00	50	149
129	OD	M	61	24,7	4,99	2,90	87,00	41	300
130	OM	M	52	33,2	3,03	9,20	102,00	46	199
131	OO	F	58	23,3	2,58	8,70	84,00	38	61
132	PTI	M	47	28,3	11,87	4,20	95,00	126	1979
133	QIK	M	53	30,3	8,06	1,45	97,00	63	984
134	QY	F	47	34,4	3,41	11,55	88,00	47	211
135	RKK	M	32	28,1	2,65	1,45	90,00	40	291
136	RMF	M	38	27,6	2,07	1,25	91,00	41	197
137	RSG	F	32	22,4	2,38	7,90	76,00	49	96
138	RY	F	70	24,4	2,01	9,15	76,50	53	276
139	SG	F	72	20,8	3,84	6,65	69,00	56	296
140	SH	M	52	29,7	6,06	14,00	104,00	57	155
141	SH	F	68	32,6	6,72	10,95	91,00	49	149
142	SH	M	41	40,8	10,19	3,40	129,00	40	234
143	SH	F	66	32,8	2,85	11,80	94,50	42	286
144	SH	M	53	25,6	3,53	2,75	90,00	33	530
145	SI	M	52	23,7	5,19	3,10	87,00	50	208
146	SK	F	60	25,4	9,30	4,40	90,00	37	212
147	SK	M	63	22,5	2,47	3,00	86,00	35	338
148	SK	M	66	26,8	9,55	2,50	96,00	33	411
149	SK	F	50	31,9	2,62	25,70	89,00	60	138
150	SM	F	65	25,5	6,05	5,10	85,00	46	146

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
151	SM	F	69	25,1	2,26	14,40	82,00	50	155
152	SM	M	66	27,2	4,29	3,90	100,00	50	410
153	SNA	F	44	24,0	2,28	2,60	78,00	31	129
154	SNF	F	51	28,2	9,14	3,25	91,00	54	261
155	SO	M	54	21,8	2,91	5,75	80,00	41	161
156	SO	M	73	28,7	2,68	5,50	94,50	26	127
157	SS	F	65	26,4	2,86	10,65	92,50	64	290
158	ST	M	54	28,9	7,90	6,70	100,00	51	576
159	SY	M	48	24,4	2,15	2,70	82,00	45	105
160	TF	M	61	24,9	5,99	5,30	91,50	67	386
161	TG	M	45	26,2	3,83	6,70	96,00	58	160
162	TGK	F	52	27,8	5,84	5,40	88,20	46	999
163	TH	M	59	34,1	7,97	4,30	108,00	45	199
164	TIM	F	47	26,7	2,43	9,55	80,00	49	171
165	TJCI	M	45	21,5	3,58	3,80	85,00	57	180
166	TK	M	61	27,3	3,11	8,70	96,25	58	167
167	TM	M	62	26,6	3,29	1,65	98,00	50	159
168	TN	F	44	35,8	10,97	2,40	95,00	50	217
169	TO	M	66	25,5	2,87	4,45	89,00	37	225
170	TOF	F	76	33,7	2,33	17,00	104,00	38	312
171	TS	M	59	23,4	2,22	4,60	82,00	36	237
172	TSY	F	51	31,1	5,37	9,75	96,00	89	1337
173	TT	M	76	26,4	2,09	5,40	94,00	49	176
174	TT	M	74	32,5	3,22	9,90	105,00	39	185
175	TT	M	46	27,4	2,97	6,15	87,00	59	266
176	TT	M	75	23,7	4,19	5,27	84,50	51	388
177	TTI	F	68	37,9	3,76	8,00	95,00	53	165
178	TTM	F	61	27,2	5,15	2,77	86,50	51	238
179	TTMM	F	48	29,7	2,97	3,10	87,00	47	122
180	TUI	F	70	21,8	2,69	8,55	75,50	52	158

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
181	TUS	F	53	25,6	2,98	8,15	84,00	45	229
182	TY	M	63	26	2,65	0,65	100,00	44	351
183	TYSJ	F	45	30,0	7,84	9,90	94,00	68	467
184	VM	M	39	32,0	5,21	3,40	104,00	42	207
185	VO	F	70	24,7	5,24	10,50	87,00	53	803
186	WTM	M	45	27,6	4,89	3,50	96,00	41	477
187	YK	M	62	26,6	2,87	3,90	94,00	67	343
188	YK	M	50	21,5	2,02	15,50	84,00	69	142
189	YK	F	68	24,4	2,54	5,45	88,00	46	260
190	YM	M	62	30,2	2,26	5,25	101,00	42	239
191	YMO	F	62	22,1	15,90	11,00	80,00	55	90
192	YN	M	61	33,6	2,65	3,25	104,00	69	1036
193	YS	F	58	27,9	11,40	6,45	86,50	44	342
194	YS	M	73	32,0	2,07	27,50	102,50	44	152
195	YT	F	69	26,0	2,91	6,00	85,00	49	251
196	YT	M	56	32,1	8,30	2,25	107,00	45	423
197	YT	F	62	25,5	2,68	4,50	76,00	46	181
198	YUM	F	35	23,1	2,39	3,00	90,00	45	413
199	YY	M	61	24,5	2,68	3,45	92,50	49	213
200	ZH	M	69	22,2	3,06	4,75	85,00	38	496

Tabela 6: Características demográficas, antropométricas e laboratoriais dos indivíduos com TGN (Artigo 3).

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
201	AB	F	64	22,9	0,93	11,45	78,00	62	149
202	AHT	F	48	18,7	0,59	19,80	66,50	42	121
203	AK	F	47	25,7	1,96	7,30	75,50	38	158
204	AMHT	F	42	22,4	1,45	9,55	74,00	44	164
205	AMO	F	47	25,5	0,93	1,70	80,50	52	204
206	AMY	M	49	21,0	1,11	14,60	79,00	48	100
207	ANO	F	65	24,3	0,62	12,15	80,50	50	76
208	AOS	F	64	22,3	1,37	1,90	80,00	44	122
209	AT	F	68	21,7	1,07	6,10	73,00	65	126
210	AT	M	62	23,1	0,80	12,85	80,50	60	----
211	ATA	M	48	22,0	0,70	3,15	80,50	50	112
212	ATFK	F	40	15,9	1,46	12,00	62,00	41	158
213	AY	F	43	27,7	1,74	8,35	87,00	70	181
214	AYS	M	41	22,4	1,03	3,10	78,50	52	132
215	CKMN	F	37	31,2	----	----	92,00	89	235
216	CMW	M	43	25,3	----	6,85	83,00	44	140
217	CNS	M	49	19,4	0,97	13,20	77,00	64	181
218	CS	F	41	22,8	-	3,2	75,50	43	137
219	CSK	M	42	18,1	1,62	19,00	74,50	37	135
220	CTTI	F	47	23,9	0,83	5,25	77,50	64	137
221	CU	M	45	30,1	1,61	2,35	97,00	35	116
222	DM	F	58	23,4	1,96	8,95	70,50	55	78
223	EEK	F	39	18,4	0,52	3,45	62,00	54	76
224	EHHT	F	41	19,5	1,99	4,60	79,50	45	70
225	EHI	M	42	22,5	0,39	11,40	80,00	50	83
226	EIG	M	51	25,3	1,72	7,00	80,00	42	231
227	EMG	M	34	27,2	1,42	3,40	85,00	43	121
228	EMI	F	56	24,4	1,86	7,00	81,00	46	132
229	ET	F	61	24,8	0,73	9,55	75,00	51	173
230	EU	F	40	20,2	0,67	4,50	78,00	44	90

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
231	EY	F	58	26,2	0,97	16,35	78,00	38	151
232	EY	F	46	28,2	1,14	4,35	83,00	40	109
233	EY	F	51	23,9	1,04	10,90	80,00	48	99
234	FHK	F	35	22,1	----	----	76,00	36	85
235	FK	F	36	24,9	1,51	8,00	78,00	51	133
236	FK	F	63	18,9	----	22,35	72,00	57	222
237	FMOO	F	42	22,0	0,25	3,65	70,00	40	94
238	FNG	F	37	20,3	0,41	19,50	76,00	48	127
239	FO	F	55	20,4	0,86	5,75	73,00	44	293
240	FO	F	63	24,3	----	5,60	92,50	66	539
241	FS	F	68	20,5	1,54	10,50	64,00	53	184
242	FS	M	62	26,5	1,58	1,00	92,00	50	257
243	FU	F	55	22,4	3,35	10,40	70,75	49	85
244	HA	M	53	23,4	1,24	1,85	81,00	56	141
245	HE	M	70	23,0	0,43	6,00	87,00	79	177
246	HH	F	44	21,0	----	12,00	70,00	48	200
247	HH	F	50	26,1	----	5,00	85,00	47	156
248	HI	F	77	22,6	1,41	8,40	79,00	49	65
249	HI	F	54	24,2	1,16	13,70	84,50	43	147
250	HI	F	51	22,5	----	18	94,00	53	57
251	HK	M	70	19,7	0,83	3,20	78,00	50	76
252	HKO	F	50	25,2	1,68	4,25	84,50	51	----
253	HKTO	F	42	20,5	1,05	24,35	65,00	56	159
254	HM	F	63	17,5	----	20,00	70,75	70	81
255	HMB	F	52	23,2	1,29	3,25	76,50	60	301
256	HMI	M	44	23,6	1,65	6,15	89,10	49	125
257	HMK	F	49	23,8	1,69	6,65	81,00	48	200
258	HMSSI	F	41	20,6	----	----	73,75	57	----
259	HN	F	74	27,4	1,81	21,70	85,00	58	188
260	HO	M	72	23,3	1,48	5,85	86,00	64	147

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
261	HO	M	60	23,0	0,85	7,15	81,00	41	124
262	HO	M	60	21,0	1,75	5,60	77,50	71	88
263	HS	F	45	21,4	0,59	6,30	74,00	61	140
264	HTG	F	59	24,3	----	2,10	80,50	72	302
265	HYA	F	49	17,2	----	10,85	53,50	77	114
266	IA	F	59	21,7	0,27	13,95	71,00	71	68
267	IK	M	43	29,3	----	----	97,00	44	164
268	IMAF	M	40	27,9	0,61	1,10	87,00	54	385
269	IMAF	F	45	22,4	1,60	10,30	71,00	48	76
270	IST	F	63	18,0	0,69	4,45	74,50	54	237
271	IYST	F	33	23,7	1,24	19,75	68,00	58	49
272	JCK	M	35	24,6	----	1,26	81,50	40	264
273	JHKC	F	47	25,0	1,55	14,75	75,00	45	162
274	JKK	F	42	24,5	0,66	13,35	78,00	49	101
275	JKN	M	45	25,5	2,45	2,25	92,00	42	277
276	JMNS	F	44	22,8	1,83	8,80	70,00	49	220
277	JTHS	F	48	23,5	----	6,35	69,50	34	197
278	JTK	M	44	25,5	0,83	9,30	80,75	54	165
279	JTT	M	41	27,0	1,32	9,28	89,00	51	153
280	KH	F	70	18,6	0,76	21,25	63,00	52	95
281	KH	F	68	23,3	1,38	13,00	83,50	51	199
282	KHK	F	54	27,5	1,82	27,85	77,50	38	165
283	KK	F	75	27,1	1,28	10,00	83,25	46	425
284	KKT	F	51	22,0	1,69	10,80	74,50	38	135
285	KKT	F	79	20,4	0,43	12,50	73,50	41	76
286	KKT	F	86	21,8	1,19	24,15	78,00	63	127
287	KM	M	46	20,5	1,85	7,35	81,50	58	165
288	KMT	F	35	17,6	----	28,5	63,00	52	92
289	KNI	F	50	21,3	0,84	8,50	66,00	56	85
290	KOY	F	65	25,8	----	3,9	87,00	42	203

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
291	KS	M	80	20,7	0,49	19,50	80,00	43	163
292	KT	M	47	23,4	0,65	6,90	81,00	105	1520
293	KY	M	67	24,0	0,56	8,85	83,00	41	100
294	LEIN	F	43	22,5	----	11,5	75,00	61	135
295	LEM	M	43	22,6	0,63	2,55	80,00	44	129
296	LH	M	52	24,0	----	6,85	85,00	42	134
297	LKO	M	67	25,9	2,41	5,20	86,50	46	483
298	LLIK	F	41	24,1	1,67	12,90	80,00	49	122
299	LMIA	F	53	27,0	1,91	11,90	87,50	58	124
300	LMOG	F	34	21,7	0,83	11,20	77,00	58	333
301	LO	M	50	25,4	0,93	5,05	82,00	55	113
302	LSY	F	44	23,6	1,39	2,50	78,00	38	114
303	LYAI	F	56	21,6	0,18	11,25	66,00	50	135
304	MAN	F	35	25,5	1,33	8,35	78,00	43	160
305	MAT	M	34	24,8	1,57	2,2	89,00	42	90
306	MF	F	75	19,2	1,31	12,06	67,00	53	167
307	MFYK	F	36	23,2	----	----	73,00	52	93
308	MH	F	50	21,7	0,74	2,50	73,00	50	219
309	MH	M	74	24,6	0,43	5,60	85,50	51	262
310	MH	F	68	23,2	----	9,3	81,10	55	149
311	MK	M	57	24,3	----	1,80	82,00	32	115
312	MK	M	40	25,2	1,29	5,35	85,00	45	141
313	MK	F	77	23,6	1,57	8,15	73,50	48	184
314	MKI	F	68	18,5	0,84	22,30	65,00	58	89
315	MKM	F	60	24,1	1,98	19,05	77,00	64	----
316	MLEHT	F	47	25,1	1,07	7,25	75,00	52	68
317	MM	F	52	23,8	1,03	3,55	74,00	42	281
318	MM	M	43	21,8	1,10	20,00	80,75	50	174
319	MM	M	51	27,8	----	----	95,00	60	579
320	MM	F	74	18,1	2,12	12,25	61,00	44	125

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
321	MMK	F	51	26,8	----	7,75	87,50	37	195
322	MMM	F	57	21,5	0,68	25,00	71,00	68	103
323	MMT	F	35	25,4	----	6,15	85,00	38	123
324	MMY	F	44	20,0	0,48	17,00	65,00	48	114
325	MO	M	64	28,2	1,95	2,15	101,75	40	140
326	MO	M	66	20,1	0,54	11,25	72,00	57	79
327	MOO	F	58	23,3	1,50	17,55	75,00	54	245
328	MOP	F	70	36,1	----	6,15	103,40	59	326
329	MS	F	59	20,7	1,20	16,95	77,25	58	149
330	MSD	F	54	22,8	1,02	5,25	73,00	53	129
331	MSE	F	48	21,8	1,35	23,65	67,25	65	67
332	MSM	F	34	21,4	1,03	4,45	68,50	53	117
333	MSY	F	69	22,9	----	6,55	78,50	55	151
334	MT	F	54	20,1	0,64	5,85	71,00	60	195
335	MT	M	66	20,8	1,54	6,30	82,00	43	223
336	MTI	F	69	25,3	1,20	22,50	85,50	65	320
337	MTSR	F	41	22,6	1,39	6,90	71,50	39	120
338	NA	F	34	22,3	0,94	4	74,00	58	77
339	NA	F	60	22,8	----	16,65	88,25	57	509
340	NF	F	41	20,2	1,56	7,85	66,75	38	164
341	NF	M	34	21,6	1,50	6,70	79,50	50	232
342	NH	M	66	28,1	1,20	3,00	93,50	40	209
343	NID	F	59	24,6	1,59	13,50	74,50	39	96
344	NK	F	52	22,7	1,66	15,70	74,00	51	88
345	NM	M	77	24,8	----	4,75	87,00	43	210
346	NM	M	45	24,6	1,97	2,40	88,00	70	110
347	NNM	F	47	22,7	0,60	26,80	72,00	41	69
348	NO	F	43	18,8	1,44	15,00	64,00	54	56
349	NO	M	69	26,2	1,07	7,80	98,50	36	133
350	NUS	F	69	26,5	1,35	16,50	83,00	44	75

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
351	OAY	M	36	22,8	0,83	6,75	81,00	45	158
352	OOF	F	62	23,9	1,58	20,65	77,00	66	94
353	OSIK	F	49	28,5	1,59	9,10	89,00	44	125
354	PMS	M	47	22,8	1,89	3,25	79,50	61	451
355									
356	RAM	M	39	23,6	0,77	7,90	86,50	52	104
357	RFU	F	57	25,1	0,88	3,7	80,50	44	223
358	RMHC	F	43	23,9	1,68	5,80	70,50	52	118
359	RNM	M	33	26,3	1,27	4,75	83,50	45	189
360	RY	F	33	17,5	----	8,1	74,00	33	124
361	SAUT	F	42	26,3	1,81	7,65	79,00	46	116
362	SE	F	51	21,9	1,03	18,50	75,25	41	66
363	SH	F	67	18,8	0,49	6,91	64,50	51	135
364	SHK	M	40	21,3	1,13	17,00	80,00	55	288
365	SIN	F	59	21,8	0,49	17,00	71,00	61	113
366	SK	F	50	22,7	2,20	6,30	77,00	37	228
367	SK	M	71	20,5	0,81	19,30	70,75	67	86
368	SKT	F	50	17,9	0,60	16,00	60,00	50	133
369	SM	M	39	24,6	2,02	4,65	79,00	45	156
370	SM	F	64	25,0	1,56	13,50	84,00	65	109
371	SM	F	62	32,3	----	11,45	101,00	47	250
372	SMA	M	44	23,4	----	----	80,00	47	234
373	SMMA	F	36	24,0	1,18	21,50	77,00	50	68
374	SN	F	54	21,6	1,54	6,35	70,00	43	111
375	SN	M	55	22,0	0,99	8,05	74,00	34	150
376	SN	M	74	24,3	0,74	20,35	94,50	57	140
377	SNM	F	49	26,9	----	15,00	80,50	42	222
378	SO	M	65	19,7	0,80	6,20	77,00	55	155
379	SO	M	51	23,1	1,54	1,6	86,50	41	318
380	SS	F	69	19,2	1,57	24,10	69,50	51	162

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
381	SSKN	F	35	20,4	1,09	5,45	64,50	60	107
382	SSN	F	80	22,7	1,07	21,00	82,25	44	130
383	SSS	F	34	22,6	1,33	----	73,00	51	205
384	SST	F	54	21,2	1,65	9,90	74,50	87	100
385	ST	F	86	22,1	0,60	12,70	87,00	62	185
386	SU	F	60	21,6	0,98	21,00	79,50	62	141
397	SWK	F	60	23,9	1,16	12,80	78,00	47	----
388	TF	M	64	19,4	1,71	16,00	76,00	60	64
389	TF	F	49	20,8	0,92	7,25	70,00	49	83
390	TFG	F	48	24,0	1,04	3,36	75,00	56	62
391	TH	M	64	18,9	0,25	14,40	71,00	55	130
392	TH	M	44	23,1	1,43	3,25	81,00	39	174
393	TH	M	40	26,2	1,75	11,75	89,00	51	596
394	TI	F	56	22,1	1,74	16,50	77,00	61	120
395	TK	F	58	19,0	0,89	18,35	67,00	50	172
396	TK	F	69	21,3	0,72	21,45	69,00	63	130
397	TK	F	81	22,1	0,92	7,88	71,50	53	134
398	TK	F	50	19,4	----	9,7	73,50	55	194
399	TK	F	75	24,1	1,89	3,18	76,00	54	108
400	TKT	F	60	24,3	1,64	5,75	86,50	58	181
401	TLNH	F	40	27,9	1,09	5,50	80,00	50	271
402	TM	M	74	21,2	0,55	9,00	76,00	35	174
403	TM	M	66	21,2	1,81	26,95	76,00	47	65
404	TM	M	67	21,6	1,09	4,30	85,15	45	164
405	TM	F	54	20,6	1,21	20,00	67,00	84	126
406	TM	F	66	31,4	----	10,5	97,00	45	108
407	TMK	F	34	20,1	0,64	6,80	64,00	48	88
408	TMKS	F	42	19,0	0,85	15,70	72,00	32	74
409	TN	M	63	21,7	----	----	80,00	34	172
410	TNH	F	65	21,8	1,45	10,7	74,00	41	117

Número	Iniciais	Sexo	Idade (anos)	IMC (kg/m ²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
411	TO	M	59	22,8	1,76	3,00	81,00	39	186
412	TS	M	85	18,3	0,33	6,70	76,00	53	177
413	TSS	M	36	17,7	0,54	15,75	67,00	46	91
414	TT	F	48	19,2	0,22	5,25	62,00	52	80
415	TTH	F	46	25,6	----	----	81,00	50	148
416	TTN	F	50	22,6	0,98	6,45	80,50	46	120
417	TTN	F	70	26,8			83,00	50	150
418	TTT	M	36	25,5	0,71	9,00	83,00	39	236
419	TTY	F	47	24,9	0,94	34,00	82,00	52	59
420	TY	F	85	20,6	0,48	14,80	79,50	55	186
421	TY	M	63	23,3	0,23	9,45	77,00	41	141
422	TY	F	70	20,4	1,06	16,65	65,50	53	455
423	VKO	M	46	27,7	----	----	92,00	48	436
424	WST	M	46	24,9	0,63	10,40	92,00	65	----
425	WSY	M	46	24,6	1,84	9,90	84,00	42	379
426	YA	M	75	24,9	1,44	5,10	87,50	50	162
427	YF	F	51	25,6	2,09	8,45	66,00	37	166
428	YI	M	36	22,6	1,45	6,20	83,75	40	63
429	YI	M	56	23,6	1,28	10,00	88,50	38	98
430	YKY	F	60	23,0	1,05	9,15	77,00	61	129
431	YLOT	F	39	24,0	1,67	6,20	75,00	50	184
432	YM	F	82	24,6	1,26	27,50	84,00	56	159
433	YM	M	59	26,4	0,72	7,55	99,00	41	213
434	YM	F	48	19,8	1,57	6,95	72,00	61	220
435	YM	F	71	23,4	----	----	76,75	59	183
436	YN	F	83	20,9	1,73	22,80	83,00	61	273
437	YO	F	71	18,2	0,65	16,00	58,00	53	98
438	YT	F	58	21,3	1,06	10,50	65,00	51	97
439	YT	M	59	25,4	----	1,45	83,50	38	183
440	YY	F	55	22,8	1,92	4,90	75,00	44	184

Tabela 7: Características demográficas, antropométricas, grau de tolerância à glicose, HOMA-R e níveis de adiponectina plasmática dos indivíduos carreadores da mutação p.Gly63ValfsX106 (Artigo 3).

Iniciais	Sexo	Idade (anos)	Toler. glicose	IMC (kg/m²)	HOMA-R	Adiponectina (µg/ml)	Cintura (cm)	HDL (mg/dl)	Triglicérides (mg/dl)
AN	F	54	DM	28,2	8,10	1,05	91,50	44	186
AN	M	56	DM	23,9	6,87	2,20	87,75	39	182
CA	M	25	TGN	20,6	0,82	0,70	67,00	51	60
MA	M	56	DM	22,9	1,25	0,60	85,00	75	759
MA	M	26	TGN	26,3	1,08	1,65	84,00	53	70
TA	M	14	TGN	22,3	1,27	0,95	68,00	37	74
TY	M	63	DM	26	2,65	0,65	100,00	44	351
YAS	F	50	DM	20,8	3,13	0,55	81,00	49	212

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